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(54) Title: A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS

#### (57) Abstract

A system for the generation of live Birnavirus such as infectious bursal disease virus (IBDV), a segmented double-stranded (ds)RNA virus of the *Birnavirdae* family, using synthetic transcripts derived from cloned DNA has been developed. Independent full-length cDNA clones were constructed which contained the entire coding and non-coding regions of RNA segments A and B of IBDV, respectively. Synthetic RNAs of both segments were produced by *in vitro* transcription of linearized plasmids with T7 RNA polymerase. Transfection of Vero cells with combined plus-sense transcripts of both segments generated infectious virus as early as 36 hours post-transfection. The development of a reverse genetics system for dsRNA viruses will greatly facilitate studies of the regulation of viral gene expression pathogenesis, and design of a new generation of live and inactivated vaccines.

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# A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS

## Background of the Invention

Infectious bursal disease virus (IBDV), a member of the *Bimaviridae* family, is the causative agent of a highly immunosuppressive disease in young chickens (Kibenge, F.S.B., et al., *J. Gen. Virol.*, 69, 1757-1775 (1988)). Infectious bursal disease (IBD) or Gumboro disease is characterized by the destruction of lymphoid follicles in the bursa of Fabricius. In a fully susceptible chicken flock of 3-6 weeks of age the clinical disease causes severe immunosuppression, and is responsible for losses due to impaired growth, decreased feed efficiency, and death. Susceptible chickens less than 3 weeks old do not exhibit outward clinical signs of the disease but have a marked infection characterized by gross lesions of the bursa.

The virus associated with the symptoms of the disease is called infectious bursal disease virus (IBDV). IBDV is a pathogen of major economic importance to the nation and world's poultry industries. It causes severe immunodeficiency in young chickens by destruction of precursors of antibody-production B cells in the bursa of Fabricius. Immunosuppression causes increased susceptibility to other diseases, and interferes with effective vaccination against Newcastle disease, Marek's disease and infectious bronchitis disease viruses.

There are two known serotypes of IBDV. Serotype I viruses are pathogenic to chickens whereas serotype II viruses infect chickens and turkeys. The infection of turkeys is presently of unknown clinical significance.

IBDV belongs to a group of viruses called *Birnaviridae* which includes other bisegmented RNA viruses such as infectious pancreatic necrosis virus (fish), tellina virus and oyster virus (bivalve mollusks) and drosophila X virus (fruit fly). These viruses all contain high molecular weight (MW) double-stranded RNA genomes.

The capsid of the IBDV virion consists of several structural proteins.

As many as nine structural proteins have been reported but there is evidence that some of these may have a precursor-product relationship (Kibenge,

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F.S.B., et al., *J. Gen. Virol.*, 69, 1757-1775 (1988)). The designation and molecular weights of the viral proteins (VP) are as shown below.

5	Viral Protein	Molecular Weight
_	VP1	90 kDa
	VP2	41 kDa
	VP3	32 kDa
	VP4	28 kDa
10	VP5	17 kDa

Two segments of double-stranded RNA were identified in the genome of IBDV. The IBDV genome consists of two segments of double-stranded (ds)RNA that vary between 2827 (segment B) to 3261 (segment A) nucleotide base pairs (Mundt, E. et al., Virology, 209, 10-18 (1995)). The larger segment A encodes a polyprotein which is cleaved by autoproteolysis to form mature viral proteins VP2, VP3 and VP4 (Hudson, P.J. et al., Nucleic Acids Res., 14, 5001-5012 (1986)). VP2 and VP3 are the major structural proteins of the virion. VP2 is the major host-protective immunogen of IBDV, and contains the antigenic regions responsible for the induction of neutralizing antibodies (Azad, et al., Virology, 161, 145-152 (1987)). A second open reading frame (ORF), preceding and partially overlapping the polyprotein gene, encodes a protein (VP5) of unknown function that is present in IBDV-infected cells (Mundt, E., et al., J. Gen. Virol., 76, 437-443, (1995)). The smaller segment B encodes VP1, a 90-kDa multifunctional protein with polymerase and capping enzyme activities (Spies, U., et al., Virus Res., 8, 127-140 (1987); Spies, U., et al., J. Gen. Virol., 71, 977-981 (1990)).

It has been demonstrated that the VP2 protein is the major host protective immunogen of IBDV, and that it contains the antigenic region responsible for the induction of neutralizing antibodies. The region containing the neutralization site has been shown to be highly conformation-dependent. The VP3 protein has been considered to be a group-specific antigen because

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it is recognized by monoclonal antibodies directed against it from strains of both serotype I and II viruses. The VP4 protein appears to be a virus-coded protease that is involved in the processing of a precursor polyprotein of the VP2, VP3 and VP4 proteins.

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Although the nucleotide sequences for genome segments A and B of various IBDV strains have been published, it was only recently that the complete 5'- and 3'-noncoding sequences of both segments were determined. The 5'-noncoding region of IBDV segments A and B contain a consensus sequence of 32 nucleotides, whereas the 3'-noncoding terminal sequences of both segments are unrelated, but conserved among IBDV strains of the same serotype (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). These terminii might contain sequences important in packaging and in the regulation of IBDV gene expression, as demonstrated for other dsRNA containing viruses such as mammalian and plant reoviruses, and rotaviruses (Anzola, et al., *Proc. Natl. Acad. Sci. USA*, 84, 8301-8305 (1987); Zou, S., et al., *Virology*, 186, 377-388 (1992); Gorziglia, M.I., et al., *Proc. Natl. Acad. Sci. USA*, 89, 5784-5788 (1992)).

In recent years, a number of infectious animal RNA viruses have been

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generated from cloned cDNA using transcripts produced by DNA-dependent RNA polymerase (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)). For example poliovirus, a plus-stranded RNA virus; influenza virus, a segmented negative-stranded RNA virus; rabies virus, a non-segmented negative-stranded RNA virus; all were recovered from cloned cDNAs of their respective genomes (van der Werf, S., et al., *Proc. Natl. Acad. Sci. USA*, 83, 2330-2334 (1986); Enami, M., et al., *Proc. Natl. Acad. Sci. USA*, 87, 3802-3805 (1990); Schnell, M.J., et al., *EMBO J.*, 13, 4195-4205 (1994)). For reovirus, it was shown that transfection of cells with a combination of SSRNA, dsRNA and *in vitro* translated reovirus products generated infectious reovirus when complemented with a helper virus from a different serotype (Roner, M.R., et al., *Virology*, 179, 845-852 (1990)). However, to date, there has been no report of a recovered infectious virus of segmented dsRNA genome from synthetic RNAs only.

## Summary of the Invention

This invention relates to the infectious bursal disease virus (IBDV) that is associated with Gumboro disease of young chickens. More particularly, this invention relates to a system for the generation of infectious bursal disease virus (IBDV) using synthetic transcripts derived from cloned cDNA. The present invention will facilitate studies of the regulation of viral gene expression, pathogenesis and design of a new generation of live and inactivated vaccines.

### Detailed Description of the Invention

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In an effort to develop a reverse genetics system for IBDV, three independent full-length cDNA clones which contain segment A of serotype I strain D78 or serotype II strain 23/82 and segment B of the serotype I strain P2, respectively, were constructed. Synthetic RNAs of segments A and B were produced by *in vitro* transcription reaction on linearized plasmids with T7 RNA polymerase. Transcripts of these segments, either untreated or treated with DNase or RNase, were evaluated for the generation of infectious virus by transfection of Vero cells.

. 20 The present inventors have demonstrated that synthetic transcripts derived from cloned DNA corresponding to the entire genome of a segmented dsRNA animal virus can give rise to a replicating virus. The recovery of infectious virus after transfecting cells with synthetic plus-sense RNAs derived from cloned cDNA of a virus with a dsRNA genome (IBDV) completes the quest of generating reverse infectious systems for RNA viruses. A number of investigators have generated infectious animal RNA viruses from cloned cDNA (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)). Van der Werf et al. were first to generate poliovirus, a plus-stranded RNA virus, using synthetic RNA produced by T7 RNA polymerase on cloned cDNA template (van der Werf, S., et al., *Proc. Natl. Acad. Sci. USA*, 83, 2330-2334 (1986)). later, Enami et al. rescued influenza virus, a segmented negative-stranded RNA virus (Enami, M., et al., *Proc. Natl. Acad. Sci. USA*, 87, 3802-3805 (1990)); and Schnell et al. generated rabies virus, a non-segmented negative-stranded RNA virus, from cloned cDNAs of their respective genomes (Schnell, M.J., et

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al., *EMBO J.*, 13, 4195-4205 (1994)). Roner et al. developed an infectious system for a segmented dsRNA reovirus by transfecting cells with a combination of synthetic ssRNA, dsRNA, *in vitro* translated reovirus products, and complemented with a helper virus of different serotype (Roner, M.R., et al., *Virology*, 179, 845-852 (1990)). The resulting virus was discriminated from the helper virus by plaque assay. However, in this system the use of a helper virus was necessary. In contrast, the presently described reverse genetics system of IBDV does not require a helper virus or other viral proteins. Transfection of cells with plus-sense RNAs of both segments was sufficient to generate infectious virus (IBDV). The fate of the additional one or four nucleotides, respectively, transcribed at the 3'-end of segment A was not determined. However, this did not prevent the replication of the viral dsRNA. Similar effects were observed for plus-stranded RNA viruses by different investigators (Boyer, J.C., et al., *Virology*, 198, 415-426 (1994)).

Transfection of plus-sense RNAs of both segments into the same cell was necessary for the successful recovery of IBDV. Transfected RNAs of both segments had to be translated by the cellular translation machinery. The polyprotein of segment A was presumably processed into VP2, VP3 and VP4 proteins which form the viral capsid. The translated protein VP1 of segment B probably acted as a RNA-dependent RNA polymerase and transcribed minus-strands from synthetic plus-strands of both segments, and the reaction products formed dsRNA. Recently, Dobos reported that in vitro transcription by the virion RNA-dependent RNA polymerase of infectious pancreatic necrosis virus (IPNV), a prototype virus of the Birnaviridae family, is primed by VP1 and then proceeds via an asymmetric, semiconservative, stranddisplacement mechanism to synthesize only plus strands during replication of the viral genome (Dobos, P., Virology, 208, 10-25 (1995)). The present system shows that synthesis of minus-strands proceeds on the plus-strands. Whether the resulting transcribed minus-strand RNA serves as a template for the transcription of plus-strands or not remains the subject of further investigation.

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To prove that the infectious IBDV contained in the supernatants of transfected cells was indeed derived from the synthetic transcripts, an artificial chimera was generated containing segment A of a serotype II strain and segment B of a serotype I strain. Sequence analysis verified this genome combination. The results also indicate that the terminal sequence motifs described by Mundt and Müller are probably responsible for replication, sorting and packaging of the viral genome (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Presence of serotype-specific terminal sequences obviously does not prevent proper replication of serotype II A segment by the action of the RNA-dependent RNA polymerase VP1 of the serotype I segment B. The ability to create recombinant viruses will greatly help in analyzing the precise function of serotype-specific and serotype-common terminal sequences.

The recovery of infectious IBDV demonstrates that only the plus-strand RNAs of both segments are sufficient to initiate replication of dsRNA. Thus, the results are in agreement with the general features of reovirus and rotavirus replication where the plus-strand RNAs serve as a template for the synthesis of progeny minus-strands to yield dsRNA (Schonberg, M., et al., *Proc. Natl. Acad. Sci.* Patton, J.T., *Virus Res.*, 6, 217-233 (1986); Chen, D., et al., *J. Virol.*, 68, 7030-7039 (1994)). However, the semiconservative, strand displacement mechanisms proposed by Spies et al. and Dobos could not be excluded (Spies, U., et al., *Virus Res.*, 8, 127-140 (1987); Dobos, P., *Virology*, 208, 10-25 (1995)). The development of a reverse genetics system for IBDV will greatly facilitate future studies of gene expression, pathogenesis, and help in the design of new generations of live and inactivated IBDV vaccines.

As used in the present application, the term "synthetic" as applied to nucleic acids indicates that it is a man made nucleic acid in contrast to a naturally occurring nucleic acid. The term implies no limitation as to the method of manufacture, which can be chemical or biological as long as the method of manufacture involves the intervention of man.

The term "cDNA" is intended to encompass any cDNA containing segments A and B and the 5' and 3' noncoding regions of segments A and B.

The term "infectious" as applied to viruses indicates that the virus has the ability to reproduce. The virus can be pathogenic or nonpathogentic and still be infectious.

The present invention provides a system for the generation of infectious bursal disease virus using synthetic RNA transcripts. This system can be used to study the regulation of viral gene expression, pathogenesis, and for the design of a new generation of live and inactivated IBDV vaccines.

The present invention provides a recombinant vector containing at least one copy of the cDNA according to the present invention. The recombinant vector may also comprise other necessary sequences such as expression control sequences, markers, amplifying genes, signal sequences, promoters, and the like, as is known in the art. Useful vectors for this purpose are plasmids, and viruses such as baculoviruses, herpes virus (HVT) and pox viruses, e.g., fowl pox virus, and the like.

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Also provided herein is a host cell transformed with the recombinant vector of the present invention or a host cell transfected with the synthetic RNA of the present invention. The host cell may be a eukaryotic or a prokaryotic host cell. Suitable examples are *E. coli*, insect cell lines such as Sf-9, chicken embryo fibroblast (CEF) cells, chicken embryo kidney (CEK) cells, African green monkey Vero cells and the like.

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Also part of this invention is an IBDV poultry vaccine comprising a poultry protecting amount of a recombinantly produced virus or portion of a virus, wherein the virus is inactivated or modified such that it is no longer virulent.

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The virus can be inactivated by chemical or physical means. Chemical inactivation can be achieved by treating the virus with, for example, enzymes, formaldehyde, β-propiolactone, ethylene-imine or a derivative thereof, an organic solvent (e.g. halogenated hydrocarbon) and or a detergent. If necessary, the inactivating substance can be neutralized after the virus has been inactivated. Physical inactivation can be carried out by subjecting the viruses to radiation such as UV light, X-radiation, or γ-radiation.

The virus can be attenuated by known methods including serial passage, deleting sequences of nucleic acids and site directed mutagenesis either before or after production of the infectious virus to produce a virus which retains sufficient antigenicity but which has reduced virulence.

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Physiologically acceptable carriers for vaccination of poultry are known in the art and need not be further described herein. In addition to being physiologically acceptable to the poultry the carrier must not interfere with the immunological response elicited by the vaccine and/or with the expression of its polypeptide product.

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Other additives, such as adjuvants and stabilizers, among others, may also be contained in the vaccine in amounts known in the art. Preferably, adjuvants such as aluminum hydroxide, aluminum phosphate, plant and animal oils, and the like, are administered with the vaccine in amounts sufficient to enhance the immune response to the IBDV. The amount of adjuvant added to the vaccine will vary depending on the nature of the adjuvant, generally ranging from about 0.1 to about 100 times the weight of the IBDV, preferably from about 1 to about 10 times the weight of the IBDV.

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The vaccine of the present invention may also contain various stabilizers. Any suitable stabilizer can be used including carbohydrates such as sorbitol, mannitol, starch, sucrose, dextrin, or glucose; proteins such as albumin or casein; and buffers such as alkaline metal phosphate and the like. A stabilizer is particularly advantageous when a dry vaccine preparation is prepared by lyophilization.

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The vaccine can be administered by any suitable known method of inoculating poultry including nasally, ophthalmically, by injection, in drinking water, in the feed, by exposure, and the like. Preferably, the vaccine is administered by mass administration techniques such as by placing the vaccine in drinking water or by spraying the animals' environment. When administered by injection, the vaccines are preferably administered parenterally. Parenteral administration as used herein means administration by intravenous, subcutaneous, intramuscular, or intraperitoneal injection.

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The vaccine of the present invention is administered to poultry to prevent IBD anytime before or after hatching. Preferably, the vaccine is administered prior to the time of birth and after the animal is about 6 weeks of age. Poultry is defined to include but not be limited to chickens, roosters, hens, broilers, roasters, breeders, layers, turkeys and ducks.

The vaccine may be provided in a sterile container in unit form or in other amounts. It is preferably stored frozen, below -20°C, and more preferably below -70°C. It is thawed prior to use, and may be refrozen immediately thereafter. For administration to poultry the recombinantly produced virus may be suspended in a carrier in an amount of about 10<sup>4</sup> to 10<sup>7</sup> pfu/ml, and more preferably about 10<sup>5</sup> to 10<sup>6</sup> pfu/ml in a carrier such as a saline solution. The inactivated vaccine may contain the antigenic equivalent of 10<sup>4</sup> to 10<sup>7</sup> pfu/ml suspended in a carrier. Other carriers may also be utilized as is known in the art. Examples of pharmaceutically acceptable carriers are diluents and inert pharmaceutical carriers known in the art. Preferably, the carrier or diluent is one compatible with the administration of the vaccine by mass administration techniques. However, the carrier or diluent may also be compatible with other administration methods such as injection, eye drops, nose drops, and the like.

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The invention also can be used to produce combination vaccines with the IBDV material. The IBDV material can be combined with antigen material of Newcastle Disease Virus Infectious Bronchitis virus, Reo virus, Adeno virus and/or the Marek virus.

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The foregoing embodiments of the present invention are further described in the following Examples. However, the present invention is not limited by the Examples, and variations will be apparent to those skilled in the art without departing from the scope of the present invention.

## Brief Description of the Drawings

Figure 1 is a schematic diagram of cDNA constructs used for synthesis of plus-sense ssRNAs of IBDV with T7 RNA polymerase. Construct pUC19FLAD78 contains the cDNA of segment A of IBDV strain D78 and the recombinant plasmid pUC18FLA23 contains the full-length cDNA of segment

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A of IBDV strain 23/82. Segment A of IBDV encodes the polyprotein (VP2-VP4-VP3), and the recently identified VP5 protein. Plasmid pUC18FLBP2 contains the cDNA of segment B of strain P2 which encodes the RNA-dependent RNA polymerase (VP1). Virus specific sequences are underlined and the T7 promoter sequences are italicized. Restriction sites are shown in boldface and identified. The cleavage sites of the linearized plasmids are shown by vertical arrows and the transcription directions are marked by horizontal arrows.

Figure 2 shows an agarose gel analysis of the transcription reaction products that were used for transfection of Vero cells. Synthetic RNAs transcribed *in vitro* using T7 RNA polymerase and linearized plasmids pUC19FLAD78 (lanes 2, 4 and 6) containing the cDNA of segment A of IBDV strain D78, and pUC18FLBP2 (lanes 1, 3 and 5) containing the cDNA of segment B of strain P2, respectively. After transcription, the reaction mixtures were either treated with DNase (lanes 1 and 2), RNase (lanes 3 and 4) or left untreated (lanes 5 and 6). Two µl of the reaction products were analyzed on 1% agarose gel. Lambda DNA, digested with *Hind* III/EcoR I, was used as markers (lane M).

Figure 3 shows a comparison of nucleotide sequences of cloned RT-PCR fragments from segments A and B of the chimeric IBDV strain 23A/P2B (bold-typed) with known sequences of segments A and B of serotype II strain 23/82 and serotype I strain P2, respectively. Nucleotide identities are marked by a colon.

Figure 4 shows the DNA sequence of pUC18FLA23.

Figure 5 shows the DNA sequence of pUC19FLAD78.

Figure 6 shows the DNA sequence of pUC18FLBP2.

#### **EXAMPLES**

Viruses and Cells. Two serotype I strains of IBDV, the attenuated P2 strain from Germany and the vaccine strain D78 (Intervet International), and one serotype II strain, the apathogenic 23/82 strain, were propagated in chicken embryo cells (CEC) and purified (Mundt, E. et al., Virology, 209, 10-18 (1995); Vakharia, V.N., et al., Virus Res., 31, 265-273 (1994)). Vero cells

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were grown in M199 medium supplemented with 5% fetal calf serum (FCS) and used for transfection experiments. Further propagation of the recovered virus and immunofluorescence studies were carried out in Vero cells (Mundt, E., et al., *J. Gen. Virol.*, 76, 437-443, (1995)). For plaque assay, monolayers of secondary CEC were prepared and used (Müller, H., et al., *Virus Res.*, 4, 297-309 (1986)).

Construction of Full-Length cDNA Clones of IBDV genome. Fulllength cDNA clones of IBDV segments A and B were independently prepared. The cDNA clones containing the entire coding region of the RNA segment A of strain D78 were prepared using standard cloning procedures and methods (Vakharia, V.N., et al., Virus Res., 31, 265-273 (1994)). By comparing the D78 terminal sequences with recently published terminal sequences of other IBDV strains (Mundt, E. et al., Virology, 209, 10-18 (1995)), it was observed that D78 cDNA clones lacked the conserved first 17 and last 10 nucleotides at the 5'- and 3'-ends, respectively. Therefore, to construct a full-length cDNA clone of segment A, two primer pairs (A5'-D78, A5-IPD78 and A3'-IPD78) were synthesized and used for PCR amplification (Table 1). The DNA segments were amplified according to the protocol of the supplier (New England Biolabs) using "Deep Vent Polymerase" (high fidelity thermophilic DNA polymerase). Amplified fragments were cloned into the EcoR I site of a pCRII vector (Invitrogen Corp.) to obtain plasmids pCRD78A5' and pCRD78A3', respectively. Each plasmid was digested with EcoR I and Sal I and the resultant fragments were ligated into EcoR I digested pUC19 to obtain plasmid pUC19FLAD78 (SEQ ID NOS:27 AND 29) which now contains a full-length cDNA copy of segment A encoding all the structural proteins (VP2, VP4 and VP3, SEQ ID NO:30) as well as the non-structural VP5 protein-(SEQ ID NO:28) (Fig. 1).

Two primer pairs (A5'-23, A5IP23 and A3'-23, A3-IP23; see Table 1) were used for reverse transcription (RT) of viral genomic dsRNA of strain 23/82 using "SuperScript RT II" (RNA directed DNA polymerase with reduced RNase H activity, GIBCO/BRL). The RT reaction products were purified by phenol/chloroform extraction and ethanol precipitation. To obtain two cDNA

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fragments bounded by primer pairs A5'-23, A5-IP23 and A3'-23, A3-IP23, respectively, RT reaction products were amplified by PCR using "Deep Vent polymerase". Both RT and PCR were carried out according to the supplier's protocol. Resulting PCR fragments were blunt-end ligated into *Sma* I cleaved pUC18 vector to obtain pUC23A5' and pUC23A3'. The 3'-end of segment A contained in plasmid pUC23A3' was ligated into the *Hind* III-*Bst*B I cleaved plasmid pUC23A5' to establish the full-length cDNA of segment A of strain 23/82. The resulting plasmid was termed pUC18FLA23 (SEQ ID NOS: 31 AND 33)(Fig. 1) and encodes structural proteins VP2, VP3 and VP4 (SEQ ID NO: 32) and non-structural protein VP5 (SEQ ID NO: 34)

To obtain cDNA clones of segment B of P2 strain, two primer pairs (B5'-P2, B5-IPP2 and B3'-P2, B3-IPP2) were designed according to the published sequences and used for RT-PCR amplification (see Table 1). Using genomic dsRNA as template, cDNA fragments were synthesized and amplified according to the supplier's protocol (Perkin-Elmer Cetus). Amplified fragments were blunt-end ligated into Sma I cleaved pBS vector (Stratagene) to obtain clones pBSP2B5' and pBSP2B3'. To construct a full-length clone of segment B, the 5'-end fragment of plasmid pBSP2B5' was first subcloned between EcoR I and Pst I sites of pUC18 vector to obtain pUCP2B5'. Then the 3'-end fragment of plasmid pBSP2B3' was inserted between the unique Bgl II and Pst I sites of plasmid pUCP2B5' to obtain a full-length plasmid pUC18FLBP2 (SEQ ID NO:25) which encodes the VP1 protein (SEQ ID NO: 26) (Fig. 1). Plasmids pUC18FLBP2, pUC18FLA23 and pUC19FLAD78 were completely sequenced by using the "Sequenase" DNA sequencing system (U.S. Biochem.), and the sequence data were analyzed using either "DNASIS" (Pharmacia) or "PC/Gene" (Intelligenetics) software. The integrity of the full-length constructs was tested by in vitro transcription and translation coupled reticulocyte lysate system using T7 RNA polymerase (Promega).

Transcription and Transfection of Synthetic RNAs. Plasmids pUC19FLAD78, pUC18FLA23 and pUC18FLBP2 were digested with *BsrG* I, *Nsi* I and *Pst* I enzymes (see Fig. 1), respectively, and used as templates for *in vitro* transcription with T7 RNA polymerase (Promega). Briefly, restriction

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enzyme cleavage assays were adjusted to 0.5% SDS and incubated with proteinase K (0.5 mg/ml) for 1 hour at 37°C. The linearized DNA templates (~3 µg) were recovered after ethanol precipitation, and were added separately to a transcription reaction mixture (50 µl) containing 40 mM Tris-HCl (pH 7.9), 10 mM NaCl, 6 mM MgCl<sub>2</sub>, 2 mM spermidine, 0.5 mM ATP, CTP and UTP each, 0.1 mM GTP, 0.25 mM cap analog [m7G(5') PPP(5') G], 120 units of "RNasin" (ribonuclease inhibitor), 150 units T7 RNA polymerase (Promega), and incubated at 37°C for 1 hour. Synthetic RNA transcripts were purified by phenol/chloroform extraction and ethanol precipitation. As controls, the transcription products were treated with either DNase or RNase (Promega) before the purification step.

Vero cells were grown to 80% confluence in 60 mm dishes and washed once with phosphate-buffered saline (PBS). Three ml of "OPTI-MEM 1" (reduced serum medium containing HEPES buffer, sodium bicarbonate, hypoxanthine, thymidine, sodium pyruvate, L-glutamine, trace elements, growth factors and phenol red; from GIBCO/BRL) were added to the monolayers, and the cells were incubated at 37°C for 1 hour in a CO2 incubator. Simultaneously, 0.15 ml of "OPTI-MEM I" was incubated with 1.25 (N-[1-(2,3-dioleyloxy)propyl]-N,N,N-"Lipofectin" reagent of μg dioleoylphosphatidylethanolamine, chloride trimethylammonium and GIBCO/BRL) for 45 min. in a polystyrene tube at room temperature. Synthetic RNA transcripts of both segments, resuspended in 0.15 ml of diethyl pyrocarbonate-treated water, were added to the OPTI-MEM-Lipofectinmixture, mixed gently, and incubated on ice for 5 min. After removing the "OPTI-MEM" from the monolayers in 60 mm dishes and replacing with fresh 1.5 ml of "OPTI-MEM", the nucleic acid containing mixture was added dropwise to the Vero cells and swirled gently. After 2 hours of incubation at 37°C, the mixture was replaced with M199 medium [CaCl<sub>2</sub> (annhydrous), Fe(NO<sub>3</sub>)<sub>3</sub> 9H<sub>2</sub>0, KCl, MgSO<sub>4</sub> (anhydrous), NaCl, NaH<sub>2</sub>PO<sub>4</sub>H<sub>2</sub>O, NaHCO<sub>3</sub>, L-Alanine, L-Arginine HCI, L-Aspartic acid, L-Cysteine HCI H,O, L-Cysteine 2HCI, L-Glutamic acid, L-Glutamine, Glycine, L-Histidine HCL H<sub>2</sub>O, L-Hydroxyproline, L-Isoleucine, L-Leucine, L-Lysine HCI, L-Methionine, L-Phenylalanine, L-

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Proline, L-Serine, L-Threonine, L-Tryptophan, L-Tyrosine 2Na 2H<sub>2</sub>O, L-Valine, Alpha tocopherol PO<sub>4</sub> Na<sub>2</sub>, Ascorbic Acid, Biotin, Calciferol, D-Calcium pantothenate, Choline chloride, Folic acid, I-Inositol, Menandione NaHSO<sub>3</sub> 3H<sub>2</sub>O, Niacin, Nicotinamide, Para-aminobenzoic acid, Pyridoxine HCl, Riboflavin, Thiamine HCl, Vitamin A Acetate, Adenine SO<sub>4</sub>, Adenylic Acid, ATP, Na<sub>2</sub>, Cholesterol, 2-Deoxy-D-Ribose, D-Glucose, Glutathione, Guanine HCl, Hypoxanthine Na, Phenol Red Na, Ribose, Sodium Acetate (anhydrous), Thymine, Tween 80, Uracil, and Xanthine Na; from Mediatech, Inc.] containing 5% FCS (without rinsing cells) and the cells were further incubated at 37°C for desired time intervals.

Identification of Generated IBDV. CEC were infected with filtered (0.2 μm) supernatant from Vero cells transfected with transcripts of pUC18FLA23 and pUC18FLP2B. 16 hours post-infection, the whole cell nucleic acids were isolated (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Primers were designed according to the published sequences and RT-PCR fragments were amplified, cloned and sequenced (Mundt, E. et al., *Virology*, 209, 10-18 (1995)). Sequence data were analyzed by using "DNASIS" software.

Immunofluorescence. Vero cells, grown on cover slips to 80% confluence, were infected with the supernatants derived from transfected Vero cells (after freeze-thawing) and incubated at 37°C for two days. The cells were then washed, fixed with acetone and treated with polyclonal rabbit anti-IBDV serum. After washing, the cells were treated with fluorescein labeled goat-anti-rabbit antibody (Kirkegaard & Perry Lab.) and examined by fluorescence microscope.

Plaque Assay. Monolayers of secondary CEC, grown in 60 mm dishes, were inoculated with the supernatants derived from transfected Vero cells. After 1 hour of infection, the cells were washed once with PBS and overlayed with 0.8% Agar noble (Difco) containing 10% tryptose phosphate broth, 2% FCS, 0.112% NaHCO<sub>3</sub>, 10<sup>3</sup> units penicillin, 10<sup>3</sup> μg/ml streptomycin, 0.25 μg/ml fungizone, 0.005% neutral red, 0.0015% phenol red. The cells

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were incubated at 37°C for 2 to 3 days until plaques could be observed and counted (Müller, H., et al., *Virus Res.*, 4, 297-309 (1986)).

Construction of Full-Length cDNA clones of IBDV Genome. To develop a reverse genetics system for the dsRNA virus IBDV, two independent cDNA clones were constructed that contain segment A of strain D78 and segment B of strain P2 (Fig. 1). Each plasmid encoded either the precursor of structural proteins (VP2, VP4, VP3) and VP5 or only VP1 protein (RNA-dependent RNA polymerase). Plasmid pUC18FLBP2 upon digestion with Pst I and transcription in vitro by T7 RNA polymerase, would yield RNA containing the correct 5'- and 3'-ends. Whereas, upon digestion with BsrG I and transcription, plasmid pUC19FLAD78 would yield RNA containing the correct 5'-end but with additional four nucleotides at the 3'end. Coupled transcription and translation of the above plasmids in a rabbit reticulocyte system yielded protein products that were correctly processed and comigrated with the marker IBDV proteins after fractionating on SDS-polyacrylamide gel and autoradiography (data not shown).

# Transcription, Transfection and Generation of Infectious Virus.

Plus-sense transcripts of IBDV segment A and B were synthesized separately *in vitro* with T7 RNA polymerase using linearized full-length cDNA plasmids as templates (see Fig. 2). Although two species of RNA transcripts were observed for segment B on a neutral gel (lanes 1 and 5), fractionation of these samples on a denaturing gel yielded only one transcript-specific band (data not shown). In order to show that plus-sense RNA transcripts of both segments are needed for the generation of infectious virus, the transcription mixtures were incubated with different nucleases, as shown in Fig. 2. Synthetic RNAs recovered after treating the transcription products with DNase (lanes 1+2), RNase (lanes 3+4) or without treatment (lanes 5+6), were used for the transfection of Vero cells. As mock control, Lipofectin alone was used. Five days post-transfection, cytopathic effect (CPE) was only visible in Vero cells transfected with combined transcripts of untreated or DNase-treated transcription products, but not with RNase-treated transcription mixtures or

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mock-transfected control. In addition, no CPE was detected when Vero cells were transfected with RNA of only segment A or B (data not shown). These results demonstrate that replication of IBDV ensued after transfection of Vero cells with plus-sense ssRNAs of both segments of IBDV. To verify that the agent causing the CPE in Vero cells was indeed IBDV, transfected Vero cells were freeze-thawed, and supernatants were clarified by centrifugation, and used to infect CEC or Vero cells. CEC infected with the supernatants derived from Vero transfected cells of untreated or DNase-treated transcription mixtures produced CPE in one day post-inoculation (Table 2). However, no CPE could be detected even after five days in CEC, with the supernatants from transfected Vero cells of RNase-treated transcription mixtures, untreated segment A or B transcription mixtures and mock-transfected Vero cells. Similarly, when Vero cells on cover slips were infected with the same supernatants as described above and examined by immunofluorescence staining after 2 days, only supernatants derived from transfected Vero cells of untreated or DNAse-treated transcription mixtures gave positive immunofluorescence signal (Table 2).

Recovery of Transfectant Virus. To determine the time point for the recovery of infectious virus, Vero cells were transfected with combined RNA transcripts of segments A and B. At 4, 8, 16, 24, 36 and 48 hours post-transfection, the supernatants were examined for the presence of transfectant virus by infectivity and plaque assays, as shown in Table 3. Our results indicate that the virus could be recovered as early as 36 hours after transfection. Virus titer was 2.3 x 10<sup>2</sup> pfu/ml which appear to drop for samples obtained later than 48 hours after transfection.

Generation of a Chimeric Virus. To prove that plus-sense ssRNA of both segments of IBDV are sufficient for recovery of infectious virus, a chimeric IBDV was generated. Plasmid pUC18FLA23 containing a full-length sequence of segment A of serotype II strain was linearized by *Nsi* I digestion and ssRNA was synthesized *in vitro* using T7 RNA polymerase. The ssRNA transcript specifies the correct 5'-end but contains one additional residue at the 3'-end (Fig. 1). Vero cells were transfected with ssRNA of segment A of

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serotype II strain 23/82 and ssRNA of segment B of serotype I strain P2. Five days after transfection when CPE was evident, the supernatant was clarified (after freeze-thawing) and used to infect CEC. After a second passage in CEC, genomic RNA of the virus was analyzed by RT-PCR and sequencing of the PCR products. Primers for segment A were deigned to specifically amplify only segment A sequences derived from the serotype II strain. Primer for segment B bound to sequences of both serotypes. The amplified fragments were cloned and sequenced. The obtained segment A sequences showed a perfect match with known segment A sequences of serotype II strain 23/82, whereas segment B sequence exhibited complete homology to published segment B sequences of serotype I strain P2 (Fig. 3).

Table 1. Oligonucleotides Used for the Construction of Full Length cDNA Clones of IBDV Genomic Segments A and B.

Nucleotide Sequence	Orientation	Name	Nucleotide Number
TAATACGACTCACTATAGGATACGATCGGTCTGACCCCGGGGGGGG	( <del>+</del> )	A5′-D78	1-31
AGAGAATTCTAATACGACTCACTATAGGATACGATCGGTCTGAC	( <del>+</del> )	A5′-23	148
TGTACAGGGGACCCGCGAACGGATCCAATT	(-)	A3'-D78	3237-3261
CGGCGAATTCATGCATAGGGGACCCGCGAACGGATC	(-)	A3′-23	3242-3261
CGTCGACTACGGGATTCTGG	(-)	A5-IPD78	1711-1730
CAGAGGCAGTACTCCGTCTG	<u> </u>	A5-IP23	1971-1990
AGTCGACGGGATTCTTGCTT	÷	A3-IPD78	1723-1742
GAAGGTGTGCGAGGAC	( <del>+</del> )	A3-IP23	1883-1900
AGAGAATTC TAATACGACTCACTATAGGATACGATGGGTCTGAC	÷	B5′-P2	1-18
CGATCTGCTGCAGGCCCCCCCCAGGCGAAGG	(-)	B3′-P2	2807-2827
CTTGAGACTCTTGTTCTCTACTCC	(-)	B5-IPP2	1915-1938
ATACAGCAAAGATCTCGGG	÷	B3-IPP2	1839-1857

Composition and location of the oligonucleotide primers used for cloning. T7 promoter sequences are marked with italic types, the virus specific sequences are underlined, and the restriction sites marked in boldface. Orientation of the virus specific sequence of the primer is shown for sense (+) and antisense (-). The positions where the primers bind (nucleotide number) are according to the published sequences of P2 strain (2).

Table 2. Generation of Infections IBDV From Synthetic RNAs of Segment A and B.

Material Transfected	CPE	Immunofluoroescence
ssRNA A+B, DNase-treated	+	+
ssRNA A+B, RNase-treated	<del>-</del>	-
ssRNA A+B, untreated	+	+
ssRNA A, untreated	-	-
ssRNA B, untreated	-	-
Lipofectin only	-	-

Vero cells were transfected with synthetic RNAs of segment A and B derived from transcription reactions that were either untreated or treated with DNase or RNase. After 5 days, the supernatants were collected, clarified by centrifugation, and analyzed for the presence of virus. The infectivity of the recovered virus was determined in CEC by the appearance of cytopathic effect (CPE) 1-2 days post-inoculation. The specificity of the recovered virus was determined by immunofluorescence staining of infected Vero cells with rabbit anti-IBDV serum.

Table 3. Recovery of Virus at Various Times Post-Transfection.

Time in hours post-transfection	CPE	Immunofluorescence	pfu/ml
4	-		0
8	· <u>-</u>	-	0
16	<del>-</del>		0
24	-	-	0
36	+	+	2.3 × 10 <sup>2</sup>
48	+	+	6.0 × 10 <sup>1</sup>

Vero cells were transfected with synthetic RNAs of segment A and B as described. The infectivity and specificity of the recovered virus was detected by CPE in CEC and immunofluorescence staining in Vero cells, respectively. Monolayers of secondary CEC were used for plaque assay after inoculating the cells with the supernatants derived from transfected Vero cells. Approximate titer of the virus was calculated as plaque forming units per ml (pfu/ml).

#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: VAKHARIA, Vikram N. MUNDT, Egbert
- (ii) TITLE OF INVENTION: A METHOD FOR GENERATING BIRNAVIRUS FROM SYNTHETIC RNA TRANSCRIPTS
  - (iii) NUMBER OF SEQUENCES: 34
    - (iv) CORRESPONDENCE ADDRESS:
      - (A) ADDRESSEE: NIKAIDO, MARMELSTEIN, MURRAY & ORAM LLP
      - (B) STREET: 655 Fifteenth Street, N. W., Suite 330 G Street Lobby
      - (C) CITY: Washington
      - (D) STATE: DC
      - (E) COUNTRY: USA
      - (F) ZIP: 20005-5701
      - (v) COMPUTER READABLE FORM:
        - (A) MEDIUM TYPE: Floppy disk
        - (B) COMPUTER: IBM PC compatible
        - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
        - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
    - (vi) CURRENT APPLICATION DATA:
      - (A) APPLICATION NUMBER: US
      - (B) FILING DATE:
      - (C) CLASSIFICATION:
  - (viii) ATTORNEY/AGENT INFORMATION:
    - (A) NAME: KITTS, Monica C.
    - (B) REGISTRATION NUMBER: 36,105
    - (C) REFERENCE/DOCKET NUMBER: P8172-6002
    - (ix) TELECOMMUNICATION INFORMATION:
      - (A) TELEPHONE: 202/638-5000
      - (B) TELEFAX: 202/638-4810
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 46 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: circular
  - (ii) MOLECULE TYPE: cDNA

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(ii) MOLECULE TYPE: cDNA
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
AATTGGATCC GTTCGCGGGT CCCCTGTACA AAGCCGAATT C 41
(2) INFORMATION FOR SEQ ID NO:3:
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 36 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: double  (D) TOPOLOGY: circular
(ii) MOLECULE TYPE: CDNA
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
CGGCGAATTC ATGCATAGGG GACCCGCGAA CGGATC 36
(2) INFORMATION FOR SEQ ID NO:4:
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 44 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: double</li> <li>(D) TOPOLOGY: circular</li> </ul>
(ii) MOLECULE TYPE: cDNA
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
GTCAGACCGA TCGTATCCTA TAGTGAGTCG TATTAGAATT CTCT 44
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(i) SEQUENCE CHARACTERISTICS:

(D) TOPOLOGY: linear

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
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(a) TYPONY TON TON GRO TO NO. C	
(2) INFORMATION FOR SEQ ID NO:6:	
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<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 120 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GGAAGCCTGA GTGAGTTGAC TGACTACAGC TACAACGGGC TGATGTCAGC CACTGCGAAC	60
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(ii) MOLECULE TYPE: DNA	· 1
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(2) INFORMATION FOR SEQ ID NO:9:	
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(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
GGAAGCCTGA GTGAACTGAC AGATGTTAGC TACAATGGGT TGATGTCTGC	AACAGCCAAC 60
ATCAACGACA AAATTGGGAA CGTCCTAGTA GGGGAAGGGG TCACCGTCCT	CAGCTTACCC 120
(2) INFORMATION FOR SEQ ID NO:10:	
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(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
TTTTCAATAG TCCACAGGCG CGAACGAAGA TCTCAGCAGC GTTCGGCATA	AAGCCTACTG 60
CTGGACAAGA CGTGGAAGAA CTCTTGATCC CCAAAGTCTG GGTGCCACCT	GAGGATCCGC 120
(2) INFORMATION FOR SEQ ID NO:11:	•
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(ii) MOLECULE TYPE: DNA

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
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CTGGACAAGA CGTGGAAGAA CTCTTGATCC CTAAAGTTTG GGTGCCACCT GAGGATCCGC	120
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(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
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(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
TAATACGACT CACTATAGGA TACGATCGGT CTGACCCCGG GGGAGTCA	48
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(ii) MOLECULE TYPE: DNA	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

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(2) INFORMATION FOR SEQ ID NO:15:	•
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(ii) MOLECULE TYPE: DNA	
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(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
CGTCGACTAC GGGATTCTGG	20
(2) INFORMATION FOR SEQ ID NO:18:	

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

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		(C)	TYPE: nuclei STRANDEDNESS TOPOLOGY: li	: single				
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	(ii)	MOLE	CULE TYPE: DN	A				
	(xi)	SEQUE	ENCE DESCRIPT	ION: SEQ I	NO:19:	ı		
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(2).	INFO	RMATIC	ON FOR SEQ ID	NO:20:				
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	(ii)	MOLE	CULE TYPE: DN	IA.				
	(xi)	SEQU	ENCE DESCRIPT	'ION: SEQ I	D NO:20:	•		
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(2)	INFO	RMATI	ON FOR SEQ II	NO:21:			 •	
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	( <u>i</u> i)	MOLE	CULE TYPE: DN	IA				
	(xi)	SEQU	ENCE DESCRIPT	TION: SEQ I	D NO:21:			

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
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<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 19 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(ii) MOLECULE TYPE: DNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
ATACAGCAAA GATCTCGGG	19
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(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 2827 base pairs  (B) TYPE: nucleic acid	

1 - 11007 1114700

- (C) STRANDEDNESS: single(D) TOPOLOGY: circular
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 112..2745

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

(X1) SEQUENCE DESCRIPTION: SEQ ID NO:25:															
GGATACGATG GGTCTGACCC TCTGGGAGTC ACGAATTAAC GTGGCTACTA GGGGCGATAC															
CCGCCGCTGG C	CCGCCGCTGG CCGCCACGTT AGTGGCTCCT CTTCTTGATG ATTCTGCCAC C ATG AGT Met Ser														
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	Pro Thr Ala G		AA GAA CTC TTG ATC CCT Lu Glu Leu Leu Ile Pro 30	213											
		lu Asp Pro Leu Al	CC AGC CCT AGT CGA CTG la Ser Pro Ser Arg Leu 15 50	261											
			TT TTG CAG CCA CGG TCT al Leu Gln Pro Arg Ser 65	309											
			AA ATA CTC CCA GAC TTA ln Ile Leu Pro Asp Leu 80	357											
			TA AAA CCC ACT CTA TCT eu Lys Pro Thr Leu Ser 95	405											
	Gly Asp Gln G		AG TAC TAC CCA ACA CAT ys Tyr Tyr Pro Thr His 110	453											
		Pro Asn Ala Tyr P	CG CCA GAC ATC GCA CTA ro Pro Asp Ile Ala Leu 25 130	501											

Leu	Lys	Gln	Met	Ile 135	Tyr	Leu	Phe	Leu	Gln 140	Val	Pro	Glu	Ala	Asn 145	GAG Glu	549
GIY	Leu	Lys	<b>Asp</b> 150	Glu	Val	Thr	Leu	Leu 155	Thr	Gln	Asn	Ile	Arg 160	Asp	AAG Lys	597
Ala	Tyr	Gly 165	AGT Ser	Gly	Thr	Tyr	Met 170	Gly	Gln	Ala	Asn	Arg 175	Leu	Val	Ala	645
Met	Lys 180	Glu	GTC Val	Ala	Thr	Gly 185	Arg	Asn	Pro	Asn	Lys 190	Asp	Pro	Leu	Lys	693
CTT Leu 195	GGG Gly	TAC Tyr	ACT Thr	TTT Phe	GAG Glu 200	AGC Ser	ATC Ile	GCG Ala	CAG Gln	CTA Leu 205	CTT Leu	GAC Asp	ATC Ile	ACA Thr	CTA Leu 210	741
CCG Pro	GTA Val	GGC Gly	CCA Pro	CCC Pro 215	GGT Gly	GAG Glu	GAT Asp	GAC Asp	AAG Lys 220	CCC Pro	TGG Trp	GTG Val	CCA Pro	CTC Leu 225	ACA Thr	789
AGA Arg	GTG Val	CCG Pro	TCA Ser 230	CGG Arg	ATG Met	TTG Leu	GTG Val	CTG Leu 235	ACG Thr	GGA Gly	GAC Asp	GTA Val	GAT Asp 240	GGC Gly	GAC Asp	837
TTT	GAG Glu	GTT Val 245	GAA Glu	GAT Asp	TAC Tyr	CTT Leu	CCC Pro 250	AAA Lys	ATC Ile	AAC Asn	CTC Leu	AAG Lys 255	TCA Ser	TCA Ser	AGT Ser	885
GGA Gly	CTA Leu 260	CCA Pro	TAT Tyr	GTA Val	GGT Gly	CGC Arg 265	ACC Thr	AAA Lys	GGA Gly	GAG Glu	ACA Thr 270	ATT Ile	GGC Gly	GAG Glu	ATG Met	933
ATA Ile 275	GCT Ala	ATC Ile	TCA Ser	AAC Asn	CAG Gln 280	TTT Phe	CTC Leu	AGA Arg	GAG Glu	CTA Leu 285	TCA Ser	ACA Thr	CTG Leu	TTG Leu	AAG Lys 290	981
CAA Gln	GGT Gly	GCA Ala	GGG Gly	ACA Thr 295	AAG Lys	GGG Gly	TCA Ser	AAC Asn	AAG Lys 300	AAG Lys	AAG Lys	CTA Leu	CTC Leu	AGC Ser 305	ATG Met	1029
TTA Leu	AGT Ser	GAC Asp	TAT Tyr 310	TGG Trp	TAC Tyr	TTA Leu	TCA Ser	TGC Cys 315	GGG Gly	CTT Leu	TTG Leu	Phe	CCA Pro 320	AAG Lys	GCT Ala	1077
GAA Glu	Arg	TAC Tyr 325	GAC Asp	AAA Lys	AGT Ser	ACA Thr	TGG Trp 330	CTC Leu	ACC Thr	AAG Lys	ACC Thr	CGG Arg 335	AAC Asn	ATA Ile	TGG Trp	1125

											ATG Met 350					1173
											ATT Ile					1221
											TTG Leu					 1269
											CTT Leu					1317
											TCA Ser				GAG Glu	1365
											CAA Gln 430				TAC Tyr	1413
Tyr 435	Ile	Leu	Thr	Arg	Gly 440	Trp	Ser	Asp	Asn	Gly 445		Pro	Met	Phe	Asn. 450	1461
Gln	Thr	Trp	Ala	Thr 455	Phe	Ala	Met	Asn	Ile 460	Ala	CCT Pro	Ala	Leu	Val 465	Val	1509
Asp	Ser	Ser	Cys 470	Leu	Ile	Met	Asn	Leu 475	Gln	Ile	AAG Lys	Thr	Tyr- 480	Gly	Gln	1557
Gly	Ser	Gly 485	Asn	Ala	Ala	Thr	Phe 490	Ile	Asn	Asn		Leu 495	Leu	Ser	Thr	1605
											CCC Pro 510				AGC Ser	1653
						Glu					ATC					1701
					Asp					Leu					CTC Leu	1749

CTT Leu	GCA Ala	CAA Gln	CCA Pro 550	GGG Gly	TAC Tyr	CTG Leu	AGT Ser	GGG Gly 555	GGG Gly	GTT Val	GAA Glu	CCA Pro	GAA Glu 560	CAA Gln	TCC Ser	1797
Ser	Pro	Thr 565	GTT Val	Glu	Leu	Asp	Leu 570	Leu	Gly	Trp	Ser	Ala 575	Thr	Tyr	Ser	1845
Lys	<b>Asp</b> 580	Leu	GGG	Ile	Tyr	Val 585	Pro	Val	Leu	Asp	Lys 590	Glu	Arg	Leu	Phe	1893
Cys 595	Ser	Ala	GCG Ala	Tyr	Pro 600	Lys	Gly	Vál	Glu	Asn 605	Lys	Ser	Leu	Lys	Ser 610	1941
AAA Lys	GTC Val	GGG	ATC Ile	GAG Glu 615	CAG Gln	GCA Ala	TAC Tyr	AAG Lys	GTA Val 620	GTC Val	AGG Arg	TAT	GAG Glu	GCG Ala 625	TTG Leu	1989
AGG Arg	TTG Leu	GTA Val	GGT Gly 630	GGT Gly	TGG Trp	AAC Asn	TAC Tyr	CCA Pro 635	CTC Leu	CTG Leu	AAC Asn	AAA Lys	GCC Ala 640	TGC	AAG Lys	2037
AAT Asn	AAC Asn	GCA Ala 645	GGC Gly	GCC Ala	GCT Ala	CGG Arg	CGG Arg 650	CAT His	CTG Leu	GAG Glu	GCC Ala	AAG Lys 655	GGG Gly	TTC Phe	CCA Pro	2085
CTC Leu	GAC Asp 660	GAG Glu	TTC Phe	CTA Leu	GCC Ala	GAG Glu 665	TGG Trp	TCT Ser	GAG Glu	CTG Leu	TCA Ser 670	GAG Glu	TTC Phe	GGT Gly	GAG Glu	2133
GCC Ala 675	TTC Phe	GAA Glu	GGC	TTC Phe	AAT Asn 680	ATC Ile	AAG Lys	CTG Leu	ACC Thr	GTA Val 685	ACA Thr	TCT Ser	GAG Glu	AGC Ser	CTA Leu 690	2181
GCC Ala	GAA Glu	CTG Leu	AAC Asn	AAG Lys 695	CCA Pro	GTA Val	CCC Pro	CCC Pro	AAG Lys 700	CCC Pro	CCA Pro	AAT Asn	GTC Val	AAC Asn 705	AGA Arg	2229
CCA Pro	GTC Val	AAC Asn	ACT Thr 710	GGG Gly	GGA Gly	CTC Leu	AAG Lys	GCA Ala 715	GTC Val	AGC Ser	ÀAC Asn	GCC Ala	CTC Leu 720	AAG Lys	ACC Thr	2277
GGT Gly	CGG <b>Ar</b> g	TAC Tyr 725	AGG Arg	AAC Asn	GAA Glu	GCC Ala	GGA Gly 730	CTG Leu	AGT Ser	GGT Gly	CTC Leu	GTC Val 735	CTT Leu	CTA Leu	GCC Ala	2325
ACA Thr	GCA Ala 740	AGA Arg	AGC Ser	CGT Arg	CTG Leu	CAA Gln 745	GAT Asp	GCA Ala	GTT Val	AAG Lys	GCC Ala 750	AAG Lys	GCA Ala	GAA Glu	GCC Ala	2373

						AAG Lys										2421
						TCA Ser										2469
						GCA Ala										2517
						GTG Val										2565
						AAC Asn 825									GCC Ala	2613
	Arg					CAG Gln									AGC Ser 850	2661
						GCC Ala										2709
						CGC							CAGC	CAT		2755
GAT	GGA1	ACC I	ACTC	AAGA	AG A	GGAC	ACTA	A TC	CCAG	ACCC	CGT	ATCC	CCG (	GCCT	CGCCT	2815
GCG	GGGG	ccc (	CC			•			:						**	2827

## (2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 878 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Ser Asp Ile Phe Asn Ser Pro Gln Ala Arg Ser Thr Ile Ser Ala 1 5 10 15

Ala Phe Gly Ile Lys Pro Thr Ala Gly Gln Asp Val Glu Glu Leu Leu

Ile Pro Lys Val Trp Val Pro Pro Glu Asp Pro Leu Ala Ser Pro Ser Arg Leu Ala Lys Phe Leu Arg Glu Asn Gly Tyr Lys Val Leu Gln Pro Arg Ser Leu Pro Glu Asn Glu Glu Tyr Glu Thr Asp Gln Ile Leu Pro Asp Leu Ala Trp Met Arg Gln Ile Glu Gly Ala Val Leu Lys Pro Thr Leu Ser Leu Pro Ile Gly Asp Gln Glu Tyr Phe Pro Lys Tyr Tyr Pro Thr His Arg Pro Ser Lys Glu Lys Pro Asn Ala Tyr Pro Pro Asp Ile Ala Leu Leu Lys Gln Met Ile Tyr Leu Phe Leu Gln Val Pro Glu Ala Asn Glu Gly Leu Lys Asp Glu Val Thr Leu Leu Thr Gln Asn Ile Arg Asp Lys Ala Tyr Gly Ser Gly Thr Tyr Met Gly Gln Ala Asn Arg Leu Val Ala Met Lys Glu Val Ala Thr Gly Arg Asn Pro Asn Lys Asp Pro Leu Lys Leu Gly Tyr Thr Phe Glu Ser Ilë Ala Gln Leu Leu Asp Ile Thr Leu Pro Val Gly Pro Pro Gly Glu Asp Asp Lys Pro Trp Val Pro Leu Thr Arg Val Pro Ser Arg Met Leu Val Leu Thr Gly Asp Val Asp Gly Asp Phe Glu Val Glu Asp Tyr Leu Pro Lys Ile Asn Leu Lys Ser Ser Ser Gly Leu Pro Tyr Val Gly Arg Thr Lys Gly Glu Thr Ile Gly Glu Met Ile Ala Ile Ser Asn Gln Phe Leu Arg Glu Leu Ser Thr Leu 

Leu Lys Gln Gly Ala Gly Thr Lys Gly Ser Asn Lys Lys Leu Leu

Ser Met Leu Ser Asp Tyr Trp Tyr Leu Ser Cys Gly Leu Leu Phe Pro Lys Ala Glu Arg Tyr Asp Lys Ser Thr Trp Leu Thr Lys Thr Arg Asn Ile Trp Ser Ala Pro Ser Pro Thr His Leu Met Ile Ser Met Ile Thr Trp Pro Val Met Ser Asn Ser Pro Asn Asn Val Leu Asn Ile Glu Gly Cys Pro Ser Leu Tyr Lys Phe Asn Pro Phe Arg Gly Gly Leu Asn Arg Ile Val Glu Trp Ile Leu Ala Pro Glu Glu Pro Lys Ala Leu Val Tyr Ala Asp Asn Ile Tyr Ile Val His Ser Asn Thr Trp Tyr Ser Ile Asp Leu Glu Lys Gly Glu Ala Asn Cys Thr Arg Gln His Met Gln Ala Ala Met Tyr Tyr Ile Leu Thr Arg Gly Trp Ser Asp Asn Gly Asp Pro Met Phe Asn Gln Thr Trp Ala Thr Phe Ala Met Asn Ile Ala Pro Ala Leu Val Val Asp Ser Ser Cys Leu Ile Met Asn Leu Gln Ile Lys Thr Tyr 465. Gly Gln Gly Ser Gly Asn Ala Ala Thr Phe Ile Asn Asn His Leu Leu Ser Thr Leu Val Leu Asp Gln Trp Asn Leu Met Arg Gln Pro Arg Pro Asp Ser Glu Glu Phe Lys Ser Ile Glu Asp Lys Leu Gly Ile Asn Phe Lys Ile Glu Arg Ser Ile Asp Asp Ile Arg Gly Lys Leu Arg Gln Leu Val Leu Leu Ala Gln Pro Gly Tyr Leu Ser Gly Gly Val Glu Pro Glu Gln Ser Ser Pro Thr Val Glu Leu Asp Leu Leu Gly Trp Ser Ala Thr

Tyr Ser Lys Asp Leu Gly Ile Tyr Val Pro Val Leu Asp Lys Glu Arq Leu Phe Cys Ser Ala Ala Tyr Pro Lys Gly Val Glu Asn Lys Ser Leu Lys Ser Lys Val Gly Ile Glu Gln Ala Tyr Lys Val Val Arg Tyr Glu Ala Leu Arg Leu Val Gly Gly Trp Asn Tyr Pro Leu Leu Asn Lys Ala Cys Lys Asn Asn Ala Gly Ala Ala Arg Arg His Leu Glu Ala Lys Gly Phe Pro Leu Asp Glu Phe Leu Ala Glu Trp Ser Glu Leu Ser Glu Phe Gly Glu Ala Phe Glu Gly Phe Asn Ile Lys Leu Thr Val Thr Ser Glu Ser Leu Ala Glu Leu Asn Lys Pro Val Pro Pro Lys Pro Pro Asn Val Asn Arg Pro Val Asn Thr Gly Gly Leu Lys Ala Val Ser Asn Ala Leu Lys Thr Gly Arg Tyr Arg Asn Glu Ala Gly Leu Ser Gly Leu Val Leu Leu Ala Thr Ala Arg Ser Arg Leu Gln Asp Ala Val Lys Ala Lys Ala Glu Ala Glu Lys Leu His Lys Ser Lys Pro Asp Asp Pro Asp Ala Asp Trp Phe Glu Arg Ser Glu Thr Leu Ser Asp Leu Leu Glu Lys Ala Asp Ile Ala Ser Lys Val Ala His Ser Ala Leu Val Glu Thr Ser Asp Ala Leu Glu Ala Val Gln Ser Thr Ser Val Tyr Thr Pro Lys Tyr Pro Glu Val Lys Asn Pro Gln Thr Ala Ser Asn Pro Val Val Gly Leu His Leu 

Pro Ala Lys Arg Ala Thr Gly Val Gln Ala Ala Leu Leu Gly Ala Gly

845 840 835 Thr Ser Arg Pro Met Gly Met Glu Ala Pro Thr Arg Ser Lys Asn Ala 860 855 850 Val Lys Met Ala Lys Arg Arg Gln Arg Gln Lys Glu Ser Arg 875 870 865 (2) INFORMATION FOR SEQ ID NO:27: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 3261 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: circular (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 97..531 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27: GGATACGATC GGTCTGACCC CGGGGGAGTC ACCCGGGGAC AGGCCGTCAA GGCCTTGTTC CAGGATGGGA CTCCTCCTTC TACAACGCTA TCATTG ATG GTT AGT AGA GAT CAG 114 Met Val Ser Arg Asp Gln 880 ACA AAC GAT CGC AGC GAT GAC AAA CCT GCA AGA TCA AAC CCA ACA GAT 162 Thr Asn Asp Arg Ser Asp Asp Lys Pro Ala Arg Ser Asn Pro Thr Asp 895 890 TGT TCC GTT CAT ACG GAG CCT TCT GAT GCC AAC AAC CGG ACC GGC GTC 210 Cys Ser Val His Thr Glu Pro Ser Asp Ala Asn Asn Arg Thr Gly Val 915 910 905 CAT TCC GGA CGA CAC CCT GGA GAA GCA CAC TCT CAG GTC AGA GAC CTC 258 His Ser Gly Arg His Pro Gly Glu Ala His Ser Gln Val Arg Asp Leu 930 925 920 GAC CTA CAA TTT GAC TGT GGG GGA CAC AGG GTC AGG GCT AAT TGT CTT 306 Asp Leu Gln Phe Asp Cys Gly Gly His Arg Val Arg Ala Asn Cys Leu 945 940 935 TTT CCC TGG ATT CCC TGG CTC AAT TGT GGG TGC TCA CTA CAC ACT GCA 354 Phe Pro Trp Ile Pro Trp Leu Asn Cys Gly Cys Ser Leu His Thr Ala

955

950

Gly Gln Trp Glu Leu Gln Val Arg Ser Asp Ala Pro Asp Cys Pro Glu 965 970 975 980	402
CCT ACC GGC CAG TTA CAA CTA CTG CAG GCT AGT GAG TCG GAG TCT CAC Pro Thr Gly Gln Leu Gln Leu Gln Ala Ser Glu Ser Glu Ser His 985 990 995	450
AGT GAG GTC AAG CAC ACT TCC TGG TGG CGT TTA TGC ACT AAA CGG CAC Ser Glu Val Lys His Thr Ser Trp Trp Arg Leu Cys Thr Lys Arg His 1000 1005 1010	<b>498</b>
CAT AAA CGC CGT GAC CTT CCA AGG AAG CCT GAG TGAACTGACA GATGTTAGCT His Lys Arg Arg Asp Leu Pro Arg Lys Pro Glu 1015 1020	551
ACAATGGGTT GATGTCTGCA ACAGCCAACA TCAACGACAA AATTGGGAAC GTCCTAGTAG	611
GGGAAGGGGT CACCGTCCTC AGCTTACCCA CATCATATGA TCTTGGGTAT GTGAGGCTTG	671
GTGACCCCAT TCCCGCAATA GGGCTTGACC CAAAAATGGT AGCCACATGT GACAGCAGTG	731
ACAGGCCCAG AGTCTACACC ATAACTGCAG CCGATGATTA CCAATTCTCA TCACAGTACC	791
AACCAGGTGG GGTAACAATC ACACTGTTCT CAGCCAACAT TGATGCCATC ACAAGCCTCA	851
GCGTTGGGGG AGAGCTCGTG TTTCAAACAA GCGTCCACGG CCTTGTACTG GGCGCCACCA	911
TCTACCTCAT AGGCTTTGAT GGGACAACGG TAATCACCAG GGCTGTGGCC GCAAACAATG	971
GGCTGACGAC CGGCACCGAC AACCTTATGC CATTCAATCT TGTGATTCCA ACAAACGAGA	1031
TAACCCAGCC AATCACATCC ATCAAACTGG AGATAGTGAC CTCCAAAAGT GGTGGTCAGG	1091
CAGGGGATCA GATGTCATGG TCGGCAAGAG GGAGCCTAGC AGTGACGATC CATGGTGGCA	1151
ACTATCCAGG GGCCCTCCGT CCCGTCACGC TAGTGGCCTA CGAAAGAGTG GCAACAGGAT	1211
CCGTCGTTAC GGTCGCTGGG GTGAGCAACT TCGAGCTGAT CCCAAATCCT GAACTAGCAA	1271
AGAACCTGGT TACAGAATAC GGCCGATTTG ACCCAGGAGC CATGAACTAC ACAAAATTGA	1331
PACTGAGTGA GAGGGACCGT CTTGGCATCA AGACCGTCTG GCCAACAAGG GAGTACACTG	1391
ACTITCGTGA ATACTICATG GAGGTGGCCG ACCICAACIC TCCCCTGAAG ATTGCAGGAG	1451
CATTCGGCTT CAAAGACATA ATCCGGGCCA TAAGGAGGAT AGCTGTGCCG GTGGTCTCCA	1511
CATTGTTCCC ACCTGCCGCT CCCCTAGCCC ATGCAATTGG GGAAGGTGTA GACTACCTGC	
GGGCGATGA GGCACAGGCT GCTTCAGGAA CTGCTCGAGC CGCGTCAGGA AAAGCAAGAG	

CTGCCTCAGG	CCGCATAAGG	CAGCTGACTC	TCGCCGCCGA	CAAGGGGTAC	GAGGTAGTCG	1691
CGAATCTATT	CCAGGTGCCC	CAGAATCCCG	TAGTCGACGG	GATTCTTGCT	TCACCTGGGG	1751
TACTCCGCGG	TGCACACAAC	CTCGACTGCG	TGTTÄÄGÄGÄ	GGGTGCCACG	CTATTCCCTG	1811
TGGTTATTAC	GACAGTGGAA	GACGCCATGA	CACCCAAAGC	ATTGAACAGC	AAAATGTTTG	1871
CTGTCATTGA	AGGCGTGCGA	GAAGACCTCC	AACCTCCATC	TCAAAGAGGA	TCCTTCATAC	1931
GAACTCTCTC	TGGACACAGA	GTCTATGGAT	ATGCTCCAGA	TGGGGTACTT	CCACTGGAGA	1991
CTGGGAGAGA	CTACACCGTT	GTCCCAATAG	ATGATGTCTG	GGACGACAGC	ATTATGCTGT	2051
CCAAAGATCC	CATACCTCCT	ATTGTGGGAA	ACAGTGGAAA	TCTAGCCATA	GCTTACATGG	2111
ATGTGTTTCG	ACCCAAAGTC	CCAATCCATG	TGGCTATGAC	GGGAGCCCTC	AATGCTTGTG	2171
GCGAGATTGA	GAAAGTAAGC	TTTAGAAGCA	CCAAGCTCGC	CACTGCACAC	CGACTTGGCC	2231
TTAGGTTGGC	TGGTCCCGGA	GCATTCGATG	TAAACACCGG	GCCCAACTGG	GCAACGTTCA	2291
TCAAACGTTT	CCCTCACAAT	CCACGCGACT	GGGACAGGCT	CCCCTACCTC	AACCTACCAT	2351
ACCTTCCACC	CAATGCAGGA	CGCCAGTACC	ACCTTGCCAT	GGCTGCATCA	GAGTTCAAAG	2411
AGACCCCCGA	ACTCGAGAGT	GCCGTCAGAG	CAATGGAAGC	AGCAGCCAAC	GTGGACCCAC	2471
TATTCCAATC	TGCACTCAGT	GTGTTCATGT	GGCTGGAAGA	GAATGGGATT	GTGACTGACA	2531
TGGCCAACTT	CGCACTCAGC	GACCCGAACG	CCCATCGGAT	GCGAAATTTT	CTTGCAAACG	2591
CACCACAAGC	AGGCAGCAAG	TCGCAAAGGG	CCAAGTACGG	GACAGCAGGC	TACGGAGTGG	2651
AGGCTCGGGG	CCCCACACCA	GAGGAAGCAC	AGAGGGAAAA	AGACACACGG	ATCTCAAAGA	2711
AGATGGAGAC	CATGGGCATC	TACTTTGCAA	CACCAGAATG	GGTAGCACTC	AATGGGCACC	2771
GAGGGCCAAG	CCCCGGCCAG	CTAAAGTACT	GGCAGAACAC	ACGAGAAATA	CCGGACCCAA	2831
ACGAGGACTA	TCTAGACTAC	GTGCATGCAG	AGAAGAGCCG	GTTGGCATCA	GAAGAACAAA	2891
TCCTAAGGGC	AGCTACGTCG	ATCTACGGGG	CTCCAGGACA	GGCAGAGCCA	CCCCAAGCTT	2,951
TCATAGACGA	AGTTGCCAAA	GTCTATGAAA	TCAACCATGG	ACGTGGCCCA	AACCAAGAAC	3011
AGATGAAAGA	TCTGCTCTTG	ACTGCGATGG	AGATGAAGCA	TCGCAATCCC	AGGCGGGCTC	.3071
TACCAAAGCO	CAAGCCAAAA	CCCAATGCTC	CAACACAGAG	ACCCCCTGGT	CGGCTGGGCC	3131
GCTGGATCAG	GACCGTCTCT	GATGAGGACC	TTGAGTGAGG	CTCCTGGGAG	TCTCCCGACA	319

CCACCCGCGC AGGTGTGGAC ACCAATTCGG CCTTACAACA TCCCAAATTG GATCCGTTCG 3251
CGGGTCCCCT 3261

- (2) INFORMATION FOR SEQ ID NO:28:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 145 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
- Met Val Ser Arg Asp Gln Thr Asn Asp Arg Ser Asp Asp Lys Pro Ala 1 5 10 15
- Arg Ser Asn Pro Thr Asp Cys Ser Val His Thr Glu Pro Ser Asp Ala
  20 25 30
- Asn Asn Arg Thr Gly Val His Ser Gly Arg His Pro Gly Glu Ala His 35 40 45
- Ser Gln Val Arg Asp Leu Asp Leu Gln Phe Asp Cys Gly Gly His Arg 50 55 60
- Val Arg Ala Asn Cys Leu Phe Pro Trp Ile Pro Trp Leu Asn Cys Gly
  65 70 75 80
- Cys Ser Leu His Thr Ala Gly Gln Trp Glu Leu Gln Val Arg Ser Asp
  85 90 95
- Ala Pro Asp Cys Pro Glu Pro Thr Gly Gln Leu Gln Leu Gln Ala 100 105 110
- Ser Glu Ser Glu Ser His Ser Glu Val Lys His Thr Ser Trp Trp Arg
- Leu Cys Thr Lys Arg His His Lys Arg Arg Asp Leu Pro Arg Lys Pro 130 135 140

Glu

- (2) INFORMATION FOR SEQ ID NO:29:
  - (i) SEQUENCE CHARACTERISTICS:

(A)	LENGTH:	3261	base	pairs
(B)	TYPE: nu	cleic	acid	1

(C) STRANDEDNESS: single

(D) TOPOLOGY: circular

### (ii) MOLECULE TYPE: cDNA

#### (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 131..3166

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

(XI) SEQUENCE DESCRIPTION: SEQ ID NO.23:	
GGATACGATC GGTCTGACCC CGGGGGAGTC ACCCGGGGAC AGGCCGTCAA GGCCTTGTTC	60
CAGGATGGGA CTCCTCCTTC TACAACGCTA TCATTGATGG TTAGTAGAGA TCAGACAAAC	120
GATCGCAGCG ATG ACA AAC CTG CAA GAT CAA ACC CAA CAG ATT GTT CCG  Met Thr Asn Leu Gln Asp Gln Thr Gln Gln Ile Val Pro  150  155	169
TTC ATA CGG AGC CTT CTG ATG CCA ACA ACC GGA CCG GCG TCC ATT CCG Phe Ile Arg Ser Leu Leu Met Pro Thr Thr Gly Pro Ala Ser Ile Pro 160 165 170	217
GAC GAC ACC CTG GAG AAG CAC ACT CTC AGG TCA GAG ACC TCG ACC TAC Asp Asp Thr Leu Glu Lys His Thr Leu Arg Ser Glu Thr Ser Thr Tyr 175 180 185 190	265
AAT TTG ACT GTG GGG GAC ACA GGG TCA GGG CTA ATT GTC TTT TTC CCT Asn Leu Thr Val Gly Asp Thr Gly Ser Gly Leu Ile Val Phe Pro 195 200 205	313
GGA TTC CCT GGC TCA ATT GTG GGT GCT CAC TAC ACA CTG CAG GGC AAT Gly Phe Pro Gly Ser Ile Val Gly Ala His Tyr Thr Leu Gln Gly Asn 210 215 220	361
GGG AAC TAC AAG TTC GAT CAG ATG CTC CTG ACT GCC CAG AAC CTA CCG Gly Asn Tyr Lys Phe Asp Gln Met Leu Leu Thr Ala Gln Asn Leu Pro 225 230 235	409
GCC AGT TAC AAC TAC TGC AGG CTA GTG AGT CGG AGT CTC ACA GTG AGG Ala Ser Tyr Asn Tyr Cys Arg Leu Val Ser Arg Ser Leu Thr Val Arg 240 245 250	457
TCA AGC ACA CTT CCT GGT GGC GTT TAT GCA CTA AAC GGC ACC ATA AAC Ser Ser Thr Leu Pro Gly Gly Val Tyr Ala Leu Asn Gly Thr Ile Asn 255 260 265 270	505
GCC GTG ACC TTC CAA GGA AGC CTG AGT GAA CTG ACA GAT GTT AGC TAC	553

Ala	Val	Thr	Phe	Gln 275	Gly	Ser	Leu	Ser	Glu 280	Leu	Thr	Asp	Val	Ser 285	Tyr	
AAT Asn	GGG Gly	TTG	ATG Met 290	TCT Ser	GCA Ala	ACA Thr	GCC Ala	AAC Asn 295	ATC Ile	AAC Asn	GAC Asp	AAA Lys	ATT Ile 300	GGG Gly	AAC Asn	601
GTC Val	CTA Leu	GTA Val 305	GGG Gly	GAA Glu	GGG Gly	GTC Val	ACC Thr 310	GTC Val	CTC Leu	AGC Ser	TTA Leu	CCC Pro 315	ACA Thr	TCA Ser	TAT Tyr	649
GAT Asp	CTT- Leu 320	GGG Gly	TAT Tyr	GTG Val	AGG Arg	CTT Leu 325	GGT Gly	GAC Asp	CCC Pro	ATT Ile	CCC Pro 330	GCA Ala	ATA Ile	GGG Gly	CTT Leu	697
GAC Asp 335	CCA Pro	AAA Lys	ATG Met	GTA Val	GCC Ala 340	ACA Thr	TGT Cys	GAC Asp	AGC Ser	AGT Ser 345	GAC Asp	AGG Arg	CCC Pro	AGA Arg	GTC Val 350	745
TAC	ACC Thr	ATA Ile	ACT Thr	GCA Ala 355	GCC Ala	GAT Asp	GAT Asp	TAC Tyr	CAA Gln 360	TTC Phe	TCA Ser	TCA Ser	CAG Gln	TAC Tyr 365	CAA Gln	793
CCA Pro	GGT Gly	GGG Gly	GTA Val 370	ACA Thr	ATC Ile	ACA Thr	Leu	TTC Phe 375	TCA Ser	GCC Ala	AAC Asn	ATT Ile	GAT Asp 380	GCC Ala	ATC Ile	841
ACA Thr	AGC Ser	CTC Leu 385	AGC Ser	GTT Val	GGG Gly	GGA Gly	GAG Glu 390	CTC Leu	GTG Val	TTT Phe	CAA Gln	ACA Thr 395	AGC Ser	GTC Val	CAC His	889
			CTG Leu							Ile		TTT Phe	GAT Asp	GGG Gly	ACA Thr	937
ACG Thr 415	GTA Val	ATC Ile	ACC Thr	AGG Arg	GCT Ala 420	Val	GCC Ala	GCA Ala	AAC Asn	AAT Asn 425	GGG Gly	CTG Leu	ACG Thr	ACC Thr	GGC Gly 430	985
			CTT Leu													1033
ACC Thr	CAG Gln	CCA Pro	ATC Ile 450	ACA Thr	TCC Ser	ATC Ile	AAA Lys	CTG Leu 455	GAG Glu	ATA Ile	GTG Val	ACC Thr	TCC Ser 460	AAA Lys	AGT Ser	1081
GGT Gly	GGT Gly	CAG Gln 465	GCA Ala	GGG Gly	GAT Asp	CAG Gln	ATG Met 470	TCA Ser	TGG Trp	TCG Ser	GCA Ala	AGA Arg 475	GGG	AGC Ser	CTA Leu	1129

GCA GTG ACG ATC CAT GGT GGC AAC TAT CCA GGG GCC CTC CGT CCC GTC 1177 Ala Val Thr Ile His Gly Gly Asn Tyr Pro Gly Ala Leu Arg Pro Val 485 ACG CTA GTG GCC TAC GAA AGA GTG GCA ACA GGA TCC GTC GTT ACG GTC 1225 Thr Leu Val Ala Tyr Glu Arg Val Ala Thr Gly Ser Val Val Thr Val 505 500 510 495 GCT GGG GTG AGC AAC TTC GAG CTG ATC CCA AAT CCT GAA CTA GCA AAG 1273 Ala Gly Val Ser Asn Phe Glu Leu Ile Pro Asn Pro Glu Leu Ala Lys 520 515 AAC CTG GTT ACA GAA TAC GGC CGA TTT GAC CCA GGA GCC ATG AAC TAC 1321 Asn Leu Val Thr Glu Tyr Gly Arg Phe Asp Pro Gly Ala Met Asn Tyr 540 · 535 530 ACA AAA TTG ATA CTG AGT GAG AGG GAC CGT CTT GGC ATC AAG ACC GTC 1369 Thr Lys Leu Ile Leu Ser Glu Arg Asp Arg Leu Gly Ile Lys Thr Val . . . . 550 545 TGG CCA ACA AGG GAG TAC ACT GAC TTT CGT GAA TAC TTC ATG GAG GTG 1417 Trp Pro Thr Arg Glu Tyr Thr Asp Phe Arg Glu Tyr Phe Met Glu Val 565 560 GCC GAC CTC AAC TCT CCC CTG AAG ATT GCA GGA GCA TTC GGC TTC AAA 1465 Ala Asp Leu Asn Ser Pro Leu Lys Ile Ala Gly Ala Phe Gly Phe Lys '585 ` 58.Q 575 GAC ATA ATC CGG GCC ATA AGG AGG ATA GCT GTG CCG GTG GTC TCC ACA 1513 Asp Ile Ile Arg Ala Ile Arg Arg Ile Ala Val Pro Val Val Ser Thr -600 595 TTG TTC CCA CCT GCC GCT CCC CTA GCC CAT GCA ATT GGG GAA GGT GTA 1561 Leu Phe Pro Pro Ala Ala Pro Leu Ala His Ala Ile Gly Glu Gly Val 615 610 GAC TAC CTG CTG GGC GAT GAG GCA CAG GCT GCT TCA GGA ACT GCT CGA 1609 Asp Tyr Leu Leu Gly Asp Glu Ala Gln Ala Ala Ser Gly Thr Ala Arg 635 630 625 GCC GCG TCA GGA AAA GCA AGA GCT GCC TCA GGC CGC ATA AGG CAG CTG 1657 Ala Ala Ser Gly Lys Ala Arg Ala Ala Ser Gly Arg Ile Arg Gln Leu 650 645 640 ACT CTC GCC GCC GAC AAG GGG TAC GAG GTA GTC GCG AAT CTA TTC CAG 1705 Thr Leu Ala Ala Asp Lys Gly Tyr Glu Val Val Ala Asn Leu Phe Gln 665 670 660 655 GTG CCC CAG AAT CCC GTA GTC GAC GGG ATT CTT GCT TCA CCT GGG GTA 1753 Val Pro Gln Asn Pro Val Val Asp Gly Ile Leu Ala Ser. Pro Gly Val 680 675

CTC Leu	CGC	GGT	GCA Ala 690	His	AAC Asn	CTC Leu	GAC Asp	TGC Cys 695	GTG Val	TTA Leu	AGA Arg	GAG Glu	GGT Gly 700	GCC Ala	ACG Thr	1801
Leu	Phe	Pro 705		Val	Ile	Thr	Thr 710	Val	Glu	Asp	Ala	Met 715	Thr	Pro	Lys	1849
GCA Ala	TTG Leu 720	AAC Asn	AGC Ser	AAA Lys	ATG Met	TTT Phe 725	GCT Ala	GTC Val	ATT	GAA Glu	GGC Gly 730	GTG Val	CGA Arg	GAA Glu	GAC Asp	1897
CTC Leu 735	CAA Gln	CCT	CCA Pro	TCT Ser	CAA Gln 740	AGA Arg	GGA Gly	TCC	TTC Phe	ATA Ile 745	CGA Arg	ACT Thr	CTC Leu	TCT Ser	GGA Gly 750	1945
CAC His	AGA Arg	GTC Val	TAT Tyr	GGA Gly 755	TAT Tyr	GCT Ala	CCA Pro	GAT Asp	GGG Gly 760	GTA Val	CTT	CCA Pro	CTG Leu	GAG Glu 765	ACT Thr	1993
GGG	AGA Arg	Asp	TAC Tyr 770	ACC Thr	GTT Val	GTC Val	CCA Pro	ATA Ile 775	GAT Asp	GAT Asp	GTC Val	TGG Trp	GAC Asp 780	GAC Asp	AGC Ser	2041
ATT Ile	ATG Met	CTG Leu 785	TCC Ser	AAA Lys	GAT Asp	CCC Pro	ATA Ile 790	CCT Pro	CCT Pro	ATT	GTG Val	GGA Gly 795	AAC Asn	AGT Ser	GGA Gly	2089
AAT Asn	CTA Leu 800	GCC Ala	ATA Ile	GCT Ala	TAC Tyr	ATG Met 805	GAT Asp	GTG Val	TTT Phe	CGA Arg	CCC Pro 810	AAA Lys	GTC Val	CCA Pro	ATC Ile	2137
CAT His 815	GTG Val	GCT Ala	ATG Met	ACG Thr	GGA Gly 820	GCC Ala	CTC Leu	AAT Asn	GCT Ala	TGT Cys 825	GGC Gly	GAG Glu	ATT Ile	GAG Glu	AAA Lys 830	2185
GTA Val	AGC Ser	TTT Phe	AGA Arg	AGC Ser 835	ACC Thr	AAG Lys	CTC Leu	GCC Ala	ACT Thr 840	GCA Ala	CAC His	CGA Arg	CTT Leu	GGC Gly 845	CTT Leu	2233
AGG Arg	TTG Leu	GCT Ala	GGT Gly 850	CCC Pro	GGA Gly	GCA Ala	TTC Phe	GAT Asp 855	GTA Val	AAC Asn	ACC Thr	GGG Gly	CČC Pro 860	AAC Asn	TGG Trp	 2281
GCA Ala	ACG Thr	TTC Phe 865	ATC Ile	AAA Lys	CGT Arg	TTC Phe	CCT Pro 870	CAC His	AAT Asn	CCA Pro	CGC Arg	GAC Asp 875	TGG Trp	GAC Asp	AGG Arg	2329
CTC Leu	CCC Pro 880	TAC Tyr	CTC Leu	AAC Asn	CTA Leu	CCA Pro 885	TAC Tyr	CTT Leu	CCA Pro	CCC Pro	AAT Asn 890	GCA Ala	GGA Gly	CGC Arg	CAG Gln	2377

											GAG Glu					2425
											AAC Asn					2473
Phe	Gln	Ser	Ala 930	Leu	Ser	Val	Phe	Met 935	Trp	Leu		Glu	Asn 940	Gly	Ile	2521
Val	Thr	Asp 945	Met	Ala	Asn	Phe	Ala 950	Leu	Ser	Asp	CCG Pro	Asn 955	Ala	His	Arg	2569
											GGC Gly 970					2617
AGG Arg 975	GCC Ala	AAG Lys	TAC Tyr	GGG Gly	ACA Thr 980	GCA Ala	GGC Gly	TAC Tyr	GGA Gly	GTG Val 985	GAG Glu	GCT Ala	CGG Arg	GGC Gly	CCC . Pro 990	2665
										Thr	CGG				Lys	.2713
				Gly					Thr		GAA Glu			Ala		2761
AAT Asn	GGG Gly	CAC His 102	Arg	GGG Gly	CCA Pro	AGC Ser	Pro	Gly	CAG Gln	CTA Leu	AAG Lys	TAC Tyr 103	Trp	CAG Gln	AAC Asn	2809
		Glu					Asn					Asp			CAT His	2857
GCÁ Ala 105	Glu	AAG Lys	AGC Ser	CGG Arg	TTG Leu 106	Ala	TCA Ser	GAA Glu	GAA Glu	CAA Gln 106	Ile	CTA Leu	AGG Arg	GCA Ala	GCT Ala 1070	2905
ACG Thr	TCG Ser	ATC Ile	TAC Tyr	GGG Gly 107	Ala	CCA Pro	GGA Gly	CAG Gln	GCA Ala 108	Glu	CCA Pro	CCC	CAA Gln	GCT Ala 108	TTC Phe 5	2953
				Ala					Ile					Gly	CCA Pro	3001

AAC Asr	CAA	GAZ Glu 110	i Gin	ATG Met	AAA Lys	GAT Asp	CTG Leu 111	Leu	TTG Leu	ACT Thr	GCG Ala	ATO Met	Glu	ATG Met	Lys	3049
CAT	CGC Arg 112	AST	CCC Pro	AGG Arg	CGG Arg	GCT Ala 112	Leu	CCA Pro	AAG Lys	CCC Pro	AAG Lys 113	Pro	AAA Lys	CCC	AAT Asn	3097
GCT Ala 113	Pro	ACA Thr	CAG Gln	AGA Arg	CCC Pro 114	Pro	GGT Gly	CGG Arg	CTG Leu	GGC Gly 114	Arg	TGG Trp	ATC	AGG Arg	ACC Thr 1150	3145
GTC Val	TCT	GAT Asp	GAG Glu	GAC Asp 115	Leu	GAG Glu	TGA	GGCT(	CCT	GGGA	GTCT(	cc c	GACA	CCAC	С	3196
CGC	GCAG	GTG	TGGA	CACC	AA T	TCGG	CCTT	A CA	ACAT	CCCA	AAT"	TGGA	TCC (	STTC	GCGGGT	3256
CCC	CT												•			3261
(2)				ENCE LEN	CHAI IGTH		RIST	rics:		is						
			(D)	TOE	POLO	3Y: 1	inea	ar								
			MOLE SEQUE						) ID	NO:3						
Met 1	Thr	Asn	Leu	Gln 5	Asp	Gln	Thr	Gln	Gln 10	Ile	Val	Pro	Phe	Ile 15	Arg	
Ser	Leu	Leu	Met 20	Pro	Thr	Thr	Gly	Pro 25	Ala	Ser	Ile	Pro	Asp 30	Asp <sub>.</sub>	Thr	
Leu	Glu	Lys 35	His	Thr	Leu	Arg	Ser 40	Glu	Thr	Ser	Thr	Tyr 45	Asn	Leu	Thr	
Val	Gly 50	Asp	Thr	Gly	Ser	Gly 55	Leu	Ile	Val	Phe	Phe 60	Pro	Gly	Phe	Pro	
Gly 65	Ser	Ile	Val	Gly	Ala 70	His	Tyr	Thr	Leu	Gln 75	Gly	Asn	Gly	Asn	Tyr 80	
Lys	Phe	Asp	Gln	Met 85	Leu	Leu	Thr	Äla	Gln 90	Asn	Leu	Pro	Ala	Ser 95	Tyr	
Asn	Tyr	Cys	Arg 100	Leu	Val	Ser .	Arg	Ser 105	Leu	Thr	Val	Arg	Ser	Ser	Thr	

- Leu Pro Gly Gly Val Tyr Ala Leu Asn Gly Thr Ile Asn Ala Val Thr 115 120 125
- Phe Gln Gly Ser Leu Ser Glu Leu Thr Asp Val Ser Tyr Asn Gly Leu 130 135 140
- Met Ser Ala Thr Ala Asn Ile Asn Asp Lys Ile Gly Asn Val Leu Val 145 150 155 160
- Gly Glu Gly Val Thr Val Leu Ser Leu Pro Thr Ser Tyr Asp Leu Gly
  165 170 175
- Tyr Val Arg Leu Gly Asp Pro Ile Pro Ala Ile Gly Leu Asp Pro Lys
  180 185 190
- Met Val Ala Thr Cys Asp Ser Ser Asp Arg Pro Arg Val Tyr Thr Ile 195 200 205
- Thr Ala Ala Asp Asp Tyr Gln Phe Ser Ser Gln Tyr Gln Pro Gly Gly 210 215 220
- Val Thr Ile Thr Leu Phe Ser Ala Asn Ile Asp Ala Ile Thr Ser Leu 225 230 235 240
- Ser Val Gly Glu Leu Val Phe Gln Thr Ser Val His Gly Leu Val 245 250 255
- Leu Gly Ala Thr Ile Tyr Leu Ile Gly Phe Asp Gly Thr Thr Val Ile 260 265 270
- Thr Arg Ala Val Ala Ala Asn Asn Gly Leu Thr Thr Gly Thr Asp Asn 275 280 285
- Leu Met Pro Phe Asn Leu Val Ile Pro Thr Asn Glu Ile Thr Gln Pro 290 295 300
- Ile Thr Ser Ile Lys Leu Glu Ile Val Thr Ser Lys Ser Gly Gln 305 310 315 320
- Ala Gly Asp Gln Met Ser Trp Ser Ala Arg Gly Ser Leu Ala Val Thr 325 330 335
- Ile His Gly Gly Asn Tyr Pro Gly Ala Leu Arg Pro Val Thr Leu Val 340 345 350
- Ala Tyr Glu Arg Val Ala Thr Gly Ser Val Val Thr Val Ala Gly Val 355 360 365
- Ser Asn Phe Glu Leu Ile Pro Asn Pro Glu Leu Ala Lys Asn Leu Val 370 375 380
- Thr Glu Tyr Gly Arg Phe Asp Pro Gly Ala Met Asn Tyr Thr Lys Leu

Ile Leu Ser Glu Arg Asp Arg Leu Gly Ile Lys Thr Val Trp Pro Thr Arg Glu Tyr Thr Asp Phe Arg Glu Tyr Phe Met Glu Val Ala Asp Leu Asn Ser Pro Leu Lys Ile Ala Gly Ala Phe Gly Phe Lys Asp Ile Ile Arg Ala Ile Arg Arg Ile Ala Val Pro Val Val Ser Thr Leu Phe Pro Pro Ala Ala Pro Leu Ala His Ala Ile Gly Glu Gly Val Asp Tyr Leu Leu Gly Asp Glu Ala Gln Ala Ser Gly Thr Ala Arg Ala Ala Ser Gly Lys Ala Arg Ala Ala Ser Gly Arg Ile Arg Gln Leu Thr Leu Ala Ala Asp Lys Gly Tyr Glu Val Val Ala Asn Leu Phe Gln Val Pro Gln Asn Pro Val Val Asp Gly Ile Leu Ala Ser Pro Gly Val Leu Arg Gly Ala His Asn Leu Asp Cys Val Leu Arg Glu Gly Ala Thr Leu Phe Pro . 555 Val Val Ile Thr Thr Val Glu Asp Ala Met Thr Pro Lys Ala Leu Asn Ser Lys Met Phe Ala Val Ile Glu Gly Val Arg Glu Asp Leu Gln Pro Pro Ser Gln Arg Gly Ser Phe Ile Arg Thr Leu Ser Gly His Arg Val Tyr Gly Tyr Ala Pro Asp Gly Val Leu Pro Leu Glu Thr Gly Arg Asp Tyr Thr Val Val Pro Ile Asp Asp Val Trp Asp Asp Ser Ile Met Leu Ser Lys Asp Pro Ile Pro Pro Ile Val Gly Asn Ser Gly Asn Leu Ala Ile Ala Tyr Met Asp Val Phe Arg Pro Lys Val Pro Ile His Val Ala 

- Met Thr Gly Ala Leu Asn Ala Cys Gly Glu Ile Glu Lys Val Ser Phe 675 680 685
- Arg Ser Thr Lys Leu Ala Thr Ala His Arg Leu Gly Leu Arg Leu Ala 690 695 700
- Gly Pro Gly Ala Phe Asp Val Asn Thr Gly Pro Asn Trp Ala Thr Phe 705 710 715 720
- Ile Lys Arg Phe Pro His Asn Pro Arg Asp Trp Asp Arg Leu Pro Tyr
  725 730 735
- Leu Asn Leu Pro Tyr Leu Pro Pro Asn Ala Gly Arg Gln Tyr His Leu 740 745 750
- Ala Met Ala Ala Ser Glu Phe Lys Glu Thr Pro Glu Leu Glu Ser Ala 755 760 765
- Val Arg Ala Met Glu Ala Ala Ala Asn Val Asp Pro Leu Phe Gln Ser 770 775 780
- Ala Leu Ser Val Phe Met Trp Leu Glu Glu Asn Gly Ile Val Thr Asp 785 790 795 800
- Met Ala Asn Phe Ala Leu Ser Asp Pro Asn Ala His Arg Met Arg Asn 805 810 815
- Phe Leu Ala Asn Ala Pro Gln Ala Gly Ser Lys Ser Gln Arg Ala Lys 820 825 830
- Tyr Gly Thr Ala Gly Tyr Gly Val Glu Ala Arg Gly Pro Thr Pro Glu 835 840 845
- Glu Ala Gln Arg Glu Lys Asp Thr Arg Ile Ser Lys Lys Met Glu Thr 850 855 860
- Met Gly Ile Tyr Phe Ala Thr Pro Glu Trp Val Ala Leu Asn Gly His 865 870 875 880
- Arg Gly Pro Ser Pro Gly Gln Leu Lys Tyr Trp Gln Asn Thr Arg Glu 885 890 895
- Ile Pro Asp Pro Asn Glu Asp Tyr Leu Asp Tyr Val His Ala Glu Lys
  900 905 910
- Ser Arg Leu Ala Ser Glu Glu Gln Ile Leu Arg Ala Ala Thr Ser Ile 915 920 925
- Tyr Gly Ala Pro Gly Gln Ala Glu Pro Pro Gln Ala Phe Ile Asp Glu 930 935 940
- Val Ala Lys Val Tyr Glu Ile Asn His Gly Arg Gly Pro Asn Gln Glu

945				-	950					955		•			960	•
Gln	Met	Lys	Asp	Leu 965	Leu	Leu	Thr	Ala	Met 970	Glu	Met	Lys	His	Arg 975	Asn	
Pro	Arg	Arg	Ala 980	Leu	Pro	Lys	Pro	Lys 985	Pro	Lys	Pro	Asn	Ala 990	Pro	Thr	-
Gln	Arg	Pro 995	Pro	Gly	Arg	Leu	Gly 1000	Arg O	Trp	Ile	Arg	Thr 1005		Ser	Asp	
Glu	Asp 1010		Glu			•		•								•
(2)	INFO	ORMAT	rion	FOR	SEQ	ID 1	10:31	l:					•	:		
	(i)			CE CI ENGTI					~~							·
				PE:					LB							
		(0	c) si	rani	EDNE	ESS:	sing	jle					٠.			
		(I	) T(	OPOLO	GY:	circ	ular	•								
•	(ii)	MOI	ECUI	LE TY	PE:	cDN#	<b>A</b>	•		•			•		•	· ·
				•												
•	(ix)	FE <i>P</i>	TURE	£::												
				ME/F	ŒY:	CDS										
		(E	3) LC	CATI	ON:	97	531							•		•
	(xi)	SEC	UENC	CE DE	SCRI	PTIC	N: 5	SEO 1	ם אכ	)•31•						
	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,							, ng 3		· · • · ·	•					
GGAI	ACGA	ATC G	GTCI	GACC	C CG	GGGG	AGTO	ACC	CCGG	GAC	AGGC	CATO	AC 1	GCCI	TGTTC	60
стсс	ידידיכיני	ממי	ייירכייו	CTTI	יר ידים	ברתכת	י מייניים מיי	TCC	كالماليات	እጥረ		3.Cm				
	1100	<i></i>	-1001	CIII		CIGI	ACIA	. ICC	3116					Asp		114
											• • • •	1015	_	nap	GIII	
מים	אאכי .	ርአጥ	ccc	7.CC	CNT	CNC	7 7 X	COM	CAM.	<b>a</b> a.	<b></b>	~-				
Thr	Agn	Asp	Ara	AGC Ser	<b>QAI</b>	ASD	Lve	Pro	Acn	GGA	TCA	CAC	CCA	ACA	GAT	162
	1020		3		riop	1025		210	rsp	GIY	1030		Pro	inr	Asp	
										٠					•	
rgt -	TCC	GTT	CAT	ACG	GAG	CCT	TCT	GAT	GCC	AAC	GAC	CGG	ACC	GGC	GTC	210
		Val	His	Thr			Ser	Asp	Ala			Arg	Thr	Gly	Val	
1035	•				1040	ļ				1045	•				1050	
CAT	TCC	GGA	CGA	CAC	CCT	GGA	GAA	GCA	CAC	ACT	CAG	GTC	CGA	AAC	CTC	258
His	Ser	Gly	Arg	His	Pro	Gly	Glu	Ala	His	Thr	Gln	Val	Arg	Asn	Leu	230
				1055					1060					1065		
GAC	TTA	CAA	CTT	GAC	TGT	AGG	GGA	TAC	AGG	GTC	AGG	ACT	ገፈ	ጥርነጥ	הואחה	306
															~ <u> </u>	200

Asp	Leu	Gln	Leu 1070	_	Сув	Arg	Gly	Tyr 1075		Val	Arg	Thr	Asn 108		Leu	
			Ile					Cys		TGC Cys	Ser		His		GCA Ala	35 <b>4</b>
		Trp					Arg					Asp			GAA Glu	402
	Ala					Leu				AGT Ser 1125	Glu					450
					Thr										Asn -	498
			AGT Ser 1150	Asp					Pro		TGAG	GTTG/	ACT	GACT.	ACAGCT	551
ACA	ACGG	GCT (	SATG	rcag(	CC A	CTGC	GAAC	A TC	AACG	ACAA	GAT	CGGG	AAC	GTTC	TAGTTG	611
GAG	AAGG	GGT (	BACTO	GTTC:	rc ac	STCT	ACCG	A CT	rcati	ATGA	CÇT'	ragt"	TAT	GTGA	GACTCG	671
GTG	ACCC	CAT (	cccc	GCAG	CA GO	GACT	CGAC	C CG	AAGT:	rgat	GGC	CACG'	rgc ·	GACA	GTAGTG	731
ACA	GACC	CAG 2	AGTC	raca(	CC A	raac	AGCT	G CAG	GATG	AATA	CCA	ATTC	rcg	TCAC	AACTCA	791
TCC	CGAG	rgg (	CGTG	AAGA	CC A	CACT	GTTC"	r cc	GCCA	ACAT	CGA'	IGCT	CTC	ACCA	GCTTCA	851
GCG'	TTGG:	rgg '	<b>r</b> gag(	CTTG:	rc T	rcag	CCAA	G TA	ACGA:	rcca	AAG	CATT	GAA	GTGG	ACGTCA	911
CCA'	TTCA	CTT (	CATT	GGGT	TT G	ACGG	GACA	G AC	GTAG	CAGT	CAA	GGCA	GTT	GCAA	CAGACT	971
TTG	GGCT	GAC 2	AACT	GGGA	CA A	ACAA	CCTT	G TG	CCAT	<b>TCAA</b>	CCT	GGTG	GTC	CCAA	CAAATG	103
AGA'	TCAC	CCA (	GCCC	ATCA	CT T	CCAT	GAAA	C TA	GAGG'	TTGT	GAC	CTAC	AAG	ATTG	GCGGCA	109:
CCG	CTGG	TGA	CCCA	TATA	CA T	GGAC.	AGTG.	A GT	GGTA	CACT	AGC	TGTG	ACG	GTGC	ACGGAG	115
GCA	ACTA	CCC '	TGGG	GCTC	TC C	GTCC	TGTC	A CC	CTGG	TGGC	CTA	TGAA	CGA	GTGG	CTGCAG	121
GAT	CTGT	TGT	CACA	GTTG	CA G	GGGT	GAGC.	A AC	TTCG.	AGCT	AAT	cccc	AAC	CCTG	AGCTTG	127
CAA	AGAA	CCT .	AGTT.	ACAG.	AG T.	ATGG	CCGC	т тт	GACC	CCGG	AGC	AATG	AAC	TACA	CCAAAC	133
TAA	TACT	GAG	TGAG.	AGAG	AT C	GTCT	AGGC	A TC	AAGA	CAGT	CTG	GCCC	ACC	AGGG	AGTACA	139
					<b>.</b>	maa:	aamm	~ ~ ~	a>ma	mc> >	C TO C	3 CCC	Cur y	2202	mmaana	145

GAGCATITGG	CITTAAGGAC	ATAATCCGAG	CCATTCGGAA	GATTGCGGTG	CCAGTGGTAT	1511
CCACACTCTT	CCCTCCAGCT	GCACCCCTAG	CACATGCAAT	CGGAGAAGGT	GTAGACTACC	1571
TCCTGGGCGA	CGAGGCCCAA	GCAGCCTCAG	GGACAGCTCG	AGCCGCGTCA	GGAAAAGCTA	1631
GAGCTGCCTC	AGGACGAATA	AGGCAGCTAA	CTCTCGCAGC	TGACAAGGGG	TGCGAGGTAG	1691
TCGCCAACAT	GTTCCAGGTG	CCCCAGAATC	CCATTGTTGA	TGGCATTCTG	GCATCCCCAG	1751
GAATCCTGCG	TGGCGCACAC	AACCTCGACT	GCGTGCTATG	GGAGGGAGCC	ACTCTTTTCC	1811
CTGTTGTCAT	TACGACACTC	GAGGATGAGC	TGACCCCCAA	GGCACTGAAC	AGCAAAATGT	1871
TTGCTGTCAT	TGAAGGTGTG	CGAGAGGACC	TCCAGCCTCC	ATCCCAACGG	GGATCCTTCA	1931
TTCGAACTCT	CTCTGGCCAT	AGAGTCTATG	GCTATGCCCC	AGACGGAGTA	CTGCCTCTGG	1991
AGACCGGGAG	AGACTACACC	GTTGTCCCAA	TTGATGATGT	GTGGGACGAT	AGCATAATGC	2051
TGTCGCAGGA	CCCCATACCT	CCAATCATAG	GGAACAGCGG	CAACCTAGCC	ATAGCATACA	2111
TGGATGTCTT	CAGGCCCAAG	GTCCCCATCC	ACGTGGCTAT	GACAGGGGCC	CTCAATGCCC	2171
GCGGTGAGAT	CGAGAGTGTT	ACGTTCCGCA	GCACCAAACT	CGCCACAGCC	CACCGACTTG	2231
GCATGAAGTT	AGCTGGTCCT	GGAGCCTATG	ACATTAATAC	AGGACCTAAC	TGGGCAACGT	2291
TCGTCAAACG	TTTCCCTCAC	AATCCCCGAG	ACTGGGACAG	GTTGCCCTAC	CTCAACCTTC	2351
CTTATCTCCC	ACCAACAGCA	GGACGTCAGT	TCCATCTAGC	CCTGGCTGCC	TCCGAGTTCA	2411
	AGAACTCGAA			•	•	2471
					ATTGTGACCG	
ACATGGCTAA	CTTCGCCCTC	AGCGACCCAA	ACGCGCATAG	GATGAAAAAC	TTCCTAGCAA	2591
ACGCACCCCA	GGCTGGAAGC	AAGTCGCAGA	GGGCCAAGTA	TGGCACGGCA	GGCTACGGAG	2651
					CGGATCTCCA	
AGAAGATGGA	AACAATGGGC	ATCTACTTCG	CGACACCGGA	ATGGGTGGCT	CTCAACGGGC	2771
ACCGAGGCCC	AAGCCCCGGC	CAACTCAAGT	ACTGGCAAAA	CACAAGAGAA	ATACCAGAGC	2831
CCAATGAGGA	CTACCCAGAC	TATGTGCACG	CGGAGAAGAG	CCGGTTGGCG	TCAGAAGAAC	2891
	GGCAGCCACG					2951
CCTTCATAGA	CGAGGTCGCC	AGGGTCTATG	AAATCAACCA	TGGGCGTGGT	CCAAACCAGG	3011

AGCAGATGAA GGACCTGCTC CTGACTGCGA TGGAGATGAA GCATCGCAAT CCCAGGCGGG 3071
CTCCACCAAA GCCAAAGCCA AAACCCAATG CTCCATCACA GAGACCCCCT GGACGGCTGG 3131
GCCGCTGGAT CAGGACGGTC TCCGACGAGG ACTTGGAGTG AGGCTCCTGG GAGTCTCCCG 3191
ACACTACCCG CGCAGGTGTG GACACCAATT CGGCCTTCTA CCATCCCAAA TTGGATCCGT 3251
TCGCGGGTCC CCT 3264

- (2) INFORMATION FOR SEQ ID NO:32:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 145 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
- Met Val Ser Arg Asp Gln Thr Asn Asp Arg Ser Asp Asp Lys Pro Asp 1 5 10 15
- Gly Ser His Pro Thr Asp Cys Ser Val His Thr Glu Pro Ser Asp Ala 20 25 30
- Thr Gln Val Arg Asn Leu Asp Leu Gln Leu Asp Cys Arg Gly Tyr Arg
  50 55 60
- Val Arg Thr Asn Cys Leu Phe Pro Trp Ile Pro Trp Phe Ser Cys Arg 65 70 75 80
- Cys Ser Leu His Thr Ala Glu Gln Trp Glu Leu Pro Ile Arg Pro Asp 85 90 95
- Ala Pro Asp Ser Ala Glu Pro Ala Cys Gln Leu Gln Leu Gln Ala 100 105 110
- Ser Glu Gln Glu Ser Asn Arg Thr Val Lys His Thr Pro Trp Arg 115 120 125
- Leu Cys Thr Lys Arg Asn His Lys Arg Ser Asp Leu Pro Arg Lys Pro 130 135 140

Glu

# (2) INFORMATION FOR SEQ ID NO:33:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3264 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: circular
- (ii) MOLECULE TYPE: cDNA

#### (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 131..3169

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GGATACGATC G	GGTCTGACCC CGGGG	GAGTC ACCCGGGGAC	AGGCCATCAC TGCCTTGTTC	60
CTGGTTGGAA C	CTCCTCTTTC TGCTC	TACTA TCGTTGATGG	TGAGTAGAGA TCAGACAAAC	120
			CAA CAG ATT GTT CCG Gln Gln Ile Val Pro 155	169
		Pro Thr Thr Gly	CCG GCG TCC ATT CCG Pro Ala Ser Ile Pro 170	217
			GAA ACC TCG ACT TAC Glu Thr Ser Thr Tyr 190	265
			ATT GTC TTT TTC CCT Ile Val Phe Phe Pro 205	313
			ACA CTG CAG AGC AGT Thr Leu Gln Ser Ser 220	361
			GCG CAG AAC CTG CCT Ala Gln Asn Leu Pro 235	409
		Leu Val Ser Arg	AGT CTA ACC GTA CGG Ser Leu Thr Val Arg 250	457

			CTC Leu				Asn			5(	05
			TTC Phe							5	53
			ATG Met 290							60	01
			GGA Gly							64	49
			TAT Tyr						CTC Leu	6	<b>97</b>
		•	TTG Leu							7.	<b>45</b>
			ACA Thr							7	93
			GTG Val 370							8	41
	Ser		AGC Ser							8	89
-			GAA Glu							9	37
			GCA Ala							9	85
			AAC Asn							10	33
			CCC Pro 450							10	81

					ACA Thr			• .	1129
					GGG Gly 490				1177
Thr					GGA Gly			٠.	1225
					AAC Asn				1273
					CCC Pro			•	1321
					CTA Leu				1369
					GAG Glu 570				1417
					GGA Gly				1465
					GTG Val				1513
					GCA Ala				1561
					GCC Ala				1609
					GGA Gly 650				1657
					GTC Val				1705

											CTG Leu					1753
											CTA Leu					1801
											GAT Asp		Leu			1849
											GAA Glu 730					1897
															TCT Ser 750	1945
											GTA Val					1993
							Val				GAT Asp				GAT Asp	2041
											ATC Ile					2089
GGC Gly	AAC Asn 800	CTA Leu	GCC Ala	ATA Ile	GCA Ala	TAC Tyr 805	ATG Met	GAT Asp	GTC Val	TTC Phe	AGG Arg 810	CCC Pro	AAG Lys	GTC Val	CCC Pro	2137.
											CGC Arg					2185
											GCC Ala					2233
				Gly										Pro	AAC Asn	2281
			Phe					Pro					Asp		GAC Asp	2329

AGG Arg	TTG Leu 880	Pro	TAC	CTC Leu	AAC Asn	CTT Leu 885	CCT Pro	TAT	CTC Leu	CCA Pro	CCA Pro 890	ACA Thr	GCA Ala	GGA Gly	CGT Arg	2377
Gln 895		His	Leu	Ala	Leu 900	Ala	Ala	Ser	Glu	Phe 905	Lys	Glu	Thr	Pro	Glu 910	2425
Leu		Asp	Ala	Val 915	Arg	Ala	Met	Asp	Ala 920	Ala	Ala	Asn	Ala	<b>Asp</b> 925	Pro	2473
Leu	TTC Phe	Arg	930	Ala	Leu	Gln	Val	Phe 935	Met	Trp	Leu	Glu	Glu 940	Asn	Gly	2521
ATT Ile	GTG Val	ACC Thr 945	GAC Asp	ATG Met	GCT Ala	AAC Asn	TTC Phe 950	GCC Ala	CTC Leu	AGC Ser	GAC Asp	CCA Pro 955	AAC Asn	GCG Ala	CAT His	2569
Arg	ATG Met 960	Lys	Asn	Phe	Leu	Ala 965	Asn	Ala	Pro	Gln	Ala 970	Gly	Ser	Lys	Ser	2617
CAG Gln 975	AGG Arg	GCC Ala	AAG Lys	TAT Tyr	GGC Gly 980	ACG Thr	GCA Ala	GGC Gly	TAC	GGA Gly 985	GTG Val	GAG Glu	GCT Ala	CGA Arg	GGC Gly 990	2665
CCC	ACA Thr	CCA Pro	GAA Glu	GAG Glu 995	GCA Ala	CAG Gln	AGG Arg	GAA Glu	AAA Lys 1000	Asp	ACA Thr	CGG Arg	ATC Ile	TCC Ser 1005	Lys	2713
AAG Lys	ATG Met	GAA Glu	ACA Thr 1010	Met	GGC	ATC Ile	TAC Tyr	TTC Phe 1015	Ala	ACA Thr	CCG Pro	GAA Glu	TGG Trp 1020	Val	GCT Ala	2761
CTC Leu	AAC Asn	GGG Gly 1025	His	CGA Arg	GGC Gly	CCA Pro	AGC Ser 1030	Pro	GGC Gly	CAA Gln	CTC Leu	AAG Lys 1035	Tyr	TGG Trp	CAA Gln	2809
AAC Asn	ACA Thr 1040	Arg	GAA Glu	ATA Ile	CCA Pro	GAG Glu 1045	Pro	AAT Asn	GAG Glu	GAC Asp	TAC Tyr 1050	Pro	GAC Asp	TAT Tyr	GTG Val	2857
CAC His 1055	GCG Ala	GAG Glu	AAG Lys	AGC Ser	CGG Arg 1060	Leu	GCG Ala	TCA Ser	GAA Glu	GAA Glu 1065	Gln	ATC Ile	CTA Leu	CGG Arg	GCA Ala 1070	2905
GCC Ala	ACG Thr	TCG Ser	ATC Ile	TAC Tyr 1075	Gly	GCT Ala	CCA Pro	GGA Gly	CAG Gln 1080	Ala	GAA Glu	CCA Pro	CCC Pro	CAG Gln 1085	Ala	2953

TTC Phe	ATA Ile	GAC Asp	GAG Glu 1090	Val	GCC Ala	AGG Arg	GTC Val	TAT Tyr 1095	Glu	ATC Ile	AAC Asn	CAT His	GGG Gly 1100	Arg	GGT Gly	3001
			Glu					Leu		CTG Leu			Met			3049
		Arg					Ala			AAG Lys		Lys				3097
	Ala					Pro				CTG Leu 1145	Gly					3145
					Asp			TGAC	GCT(	CCT G	GGAG	STCTO	CC CG	GACAC	CTACC	3199
CGC	CAGO	TG T	rggao	CACC	AA T	rcgg	CCTT	C TAC	CCAT	CCCA	AATI	rgga:	rcc d	TTC	CGGGI	3259
CCC	CT.	•			•											3264
(2)			SEQUI (A) (B)	ENCE LEI	SEQ CHAI NGTH PE: 6	RACT: : 10:	ERIS' 13 au o ac:	rics mino id		ds		•				
	(:	ii) 1	MOLE	CULE	TYP	E: p	rote	in							-	
•	. (:	ki) :	SEQUI	ENCE	DES	CRIP'	TION	: SE	Q ID	NO:	34:			•		
Met 1	Thr	Asn	Leu	Met 5	`Asp	His	Thr	Gln	Gln 10	Ile	Val	Pro	Phe	Ile 15	Arg	
Ser	Leu	Leu	Met 20	Pro	Thr	Thr	Gly	Pro 25		Ser	Ile	Pro	Asp 30	Asp	Thr	
Leu	Glu	Lys 35	His	Thr	Leu	Arg	Ser 40		Thr	Ser	Thr	Tyr 45	Asn	Leu	Thr	
Val	Gly 50	Asp	Thr	Gly	Ser	Gly 55		Ile	Val	Phe	Phe 60		Gly	Phe	Pro	
Gly 65		Val	Val	Gly	Ala 70		Tyr	Thr	Leu	Gln 75		Ser	Gly	Asn	Tyr 80	
Gln	Phe	Asp	Gln	Met	Leu	Leu	Thr	Ala	Gln	Asn	Leu	Pro	Ala	Ser	Tyr	

Asn Tyr Cys Arg Leu Val Ser Arg Ser Leu Thr Val Arg Ser Ser Thr Leu Pro Gly Gly Val Tyr Ala Leu Asn Gly Thr Ile Asn Ala Val Thr Phe His Gly Ser Leu Ser Glu Leu Thr Asp Tyr Ser Tyr Asn Gly Leu Met Ser Ala Thr Ala Asn Ile Asn Asp Lys Ile Gly Asn Val Leu Val Gly Glu Gly Val Thr Val Leu Ser Leu Pro Thr Ser Tyr Asp Leu Ser Tyr Val Arg Leu Gly Asp Pro Ile Pro Ala Ala Gly Leu Asp Pro Lys Leu Met Ala Thr Cys Asp Ser Ser Asp Arg Pro Arg Val Tyr Thr Ile Thr Ala Ala Asp Glu Tyr Gln Phe Ser Ser Gln Leu Ile Pro Ser Gly 

Val Lys Thr Thr Leu Phe Ser Ala Asn Ile Asp Ala Leu Thr Ser Phe 

Ser Val Gly Glu Leu Val Phe Ser Gln Val Thr Ile Gln Ser Ile 

Glu Val Asp Val Thr Ile His Phe Ile Gly Phe Asp Gly Thr Asp Val 

Ala Val Lys Ala Val Ala Thr Asp Phe Gly Leu Thr Thr Gly Thr Asn 

Asn Leu Val Pro Phe Asn Leu Val Val Pro Thr Asn Glu Ile Thr Gln 

Pro Ile Thr Ser Met Lys Leu Glu Val Val Thr Tyr Lys Ile Gly Gly 

Thr Ala Gly Asp Pro Ile Ser Trp Thr Val Ser Gly Thr Leu Ala Val 

Thr Val His Gly Gly Asn Tyr Pro Gly Ala Leu Arg Pro Val Thr Leu - 345 

Val Ala Tyr Glu Arg Val Ala Ala Gly Ser Val Val Thr Val Ala Gly 

- Val Ser Asn Phe Glu Leu Ile Pro Asn Pro Glu Leu Ala Lys Asn Leu 370 375 380
- Val Thr Glu Tyr Gly Arg Phe Asp Pro Gly Ala Met Asn Tyr Thr Lys 385 390 395 400
- Leu Ile Leu Ser Glu Arg Asp Arg Leu Gly Ile Lys Thr Val Trp Pro 405 410 415
- Thr Arg Glu Tyr Thr Asp Phe Arg Glu Tyr Phe Met Glu Val Ala Asp 420 425 430
- Leu Asn Ser Pro Leu Lys Ile Ala Gly Ala Phe Gly Phe Lys Asp Ile
  435
  440
  445
- Ile Arg Ala Ile Arg Lys Ile Ala Val Pro Val Val Ser Thr Leu Phe 450 455 460
- Pro Pro Ala Ala Pro Leu Ala His Ala Ile Gly Glu Gly Val Asp Tyr 465 470 475 480
- Leu Leu Gly Asp Glu Ala Gln Ala Ala Ser Gly Thr Ala Arg Ala Ala 485 490 495
- Ser Gly Lys Ala Arg Ala Ala Ser Gly Arg Ile Arg Gln Leu Thr Leu 500 505 510
- Ala Ala Asp Lys Gly Cys Glu Val Val Ala Asn Met Phe Gln Val Pro 515 520 525
- Gln Asn Pro Ile Val Asp Gly Ile Leu Ala Ser Pro Gly Ile Leu Arg
  530 · 540
- Gly Ala His Asn Leu Asp Cys Val Leu Trp Glu Gly Ala Thr Leu Phe 545 550 555 560
- Pro Val Val Ile Thr Thr Leu Glu Asp Glu Leu Thr Pro Lys Ala Leu 565 570 575
- Asn Ser Lys Met Phe Ala Val Ile Glu Gly Val Arg Glu Asp Leu Gln 580 585 590
- Pro Pro Ser Gln Arg Gly Ser Phe Ile Arg Thr Leu Ser Gly His Arg
  595 600 605
- Val Tyr Gly Tyr Ala Pro Asp Gly Val Leu Pro Leu Glu Thr Gly Arg 610 620
- Asp Tyr Thr Val Val Pro Ile Asp Asp Val Trp Asp Asp Ser Ile Met 625 630 635

- Leu Ser Gln Asp Pro Ile Pro Pro Ile Ile Gly Asn Ser Gly Asn Leu 645 650 655
- Ala Ile Ala Tyr Met Asp Val Phe Arg Pro Lys Val Pro Ile His Val 660 665 670
- Ala Met Thr Gly Ala Leu Asn Ala Arg Gly Glu Ile Glu Ser Val Thr 675 680 685
- Phe Arg Ser Thr Lys Leu Ala Thr Ala His Arg Leu Gly Met Lys Leu 690 695 700
- Ala Gly Pro Gly Ala Tyr Asp Ile Asn Thr Gly Pro Asn Trp Ala Thr 705 710 715 720
- Phe Val Lys Arg Phe Pro His Asn Pro Arg Asp Trp Asp Arg Leu Pro 725 730 735
- Tyr Leu Asn Leu Pro Tyr Leu Pro Pro Thr Ala Gly Arg Gln Phe His
  740 745 750
- Leu Ala Leu Ala Ala Ser Glu Phe Lys Glu Thr Pro Glu Leu Glu Asp
  755 760 765
- Ala Val Arg Ala Met Asp Ala Ala Ala Asn Ala Asp Pro Leu Phe Arg 770 775 780
- Ser Ala Leu Gln Val Phe Met Trp Leu Glu Glu Asn Gly Ile Val Thr 785 790 795 800
- Asp Met Ala Asn Phe Ala Leu Ser Asp Pro Asn Ala His Arg Met Lys 805 810 815
- Asn Phe Leu Ala Asn Ala Pro Gln Ala Gly Ser Lys Ser Gln Arg Ala 820 825 830
- Lys Tyr Gly Thr Ala Gly Tyr Gly Val Glu Ala Arg Gly Pro Thr Pro 835 840 845
- Glu Glu Ala Gln Arg Glu Lys Asp Thr Arg Ile Ser Lys Lys Met Glu 850 855 860
- Thr Met Gly Ile Tyr Phe Ala Thr Pro Glu Trp Val Ala Leu Asn Gly 865 870 875 880
- His Arg Gly Pro Ser Pro Gly Gln Leu Lys Tyr Trp Gln Asn Thr Arg 885 890 895
- Glu Ile Pro Glu Pro Asn Glu Asp Tyr Pro Asp Tyr Val His Ala Glu 900 905 910

- Lys Ser Arg Leu Ala Ser Glu Glu Gln Ile Leu Arg Ala Ala Thr Ser 915 920 925
- Ile Tyr Gly Ala Pro Gly Gln Ala Glu Pro Pro Gln Ala Phe Ile Asp 930 935 940
- Glu Val Ala Arg Val Tyr Glu Ile Asn His Gly Arg Gly Pro Asn Gln 945 950 955 960
- Glu Gln Met Lys Asp Leu Leu Leu Thr Ala Met Glu Met Lys His Arg 965 970 975
- Asn Pro Arg Ala Pro Pro Lys Pro Lys Pro Lys Pro Asn Ala Pro 980 985 990
- Ser Gln Arg Pro Pro Gly Arg Leu Gly Arg Trp Ile Arg Thr Val Ser 995 1000 1005

Asp Glu Asp Leu Glu 1010

#### Claims

1. A method for preparing live Birnavirus, comprising the following steps:

preparing a cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce synthetic RNA transcripts, transfecting host cells with said synthetic RNA transcripts, incubating said host cells in a culture medium, and isolating live infectious bursal disease virus from said culture medium.

- 2. The method according to claim 1, wherein said Birnavirus is infectious bursal disease virus.
- 3. The method according to claim 1, wherein said host cells are African green monkey Vero cells.
- 4. The method according to claim 1, wherein said segments A and B of said cDNA are independently prepared.
- 5. The method according to claim 4, wherein said segment A is present in plasmid pUC19FLAD78 or pUC18FLA23.
- 6. The method according to claim 4, wherein said segment B is present in plasmid pUC18FLBP2.
- 7. A live infectious bursal disease virus, wherein said virus is made by a process comprising the steps of preparing a cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce a synthetic RNA transcript, transfecting a host cell with said synthetic RNA transcript, incubating said host cell in a culture medium, and isolating live infectious bursal disease virus from said culture medium.

- 8. A synthetic RNA encoding proteins VP1, VP2, VP3, VP4, and VP5 of infectious bursal disease virus.
  - 9. A host cell transfected with the synthetic RNA according to claim 8.
- 10. A cDNA containing at least a portion of the infectious bursal disease virus genome selected from the group consisting of segment A,

segment B and segments A and B of infectious bursal disease virus, wherein said cDNA includes the 5' and 3' terminii of said segments.

- 11. A recombinant vector comprising the cDNA according to claim 10.
- 12. The vector according to claim 11, wherein said vector is a plasmid.
- 13. The vector according to claim 12, wherein said plasmid is selected from the group consisting of pUC19FLAD78, pUC18FLA23 and pUC19FLBP2.
  - 14. A host cell transformed with the vector according to claim 11.
- 15. A vaccine comprising an infectious bursal disease virus according to claim 7, wherein said infectious bursal disease virus is inactivated or attenuated prior to administration.
- 16. A method for producing a live infectious bursal disease virus vaccine, comprising the steps of

preparing a full-length cDNA containing infectious bursal disease virus genome segments A and B,

transcribing said cDNA to produce synthetic RNA transcripts,
purifying said synthetic RNA transcripts,
transfecting host cells with said purified RNA transcripts,
incubating said host cells in a culture medium,
isolating live infectious bursal disease virus from said culture medium,
attenuating said live infectious bursal disease virus to produce a virus
with reduced virulence, and

combining said live infectious bursal disease virus with a pharmaceutically acceptable carrier to produce a live infectious bursal disease virus vaccine.

- 17. The method according to claim 16, wherein said live infectious bursal disease virus is attenuated by serial passage or site directed mutagenesis.
- 18. The method according to claim 1, wherein said host cells are poultry cells.
- 19. The method according to claim 18, wherein said poultry cells are chicken, turkey, or quail cells.

20. The method according to claim 19, wherein said poultry cells are chicken embryo fibroblast cells or chicken embryo kidney cells.

Fig. 1

Fig. 1A

Fig. IB

Fig. IC

Fig. 4

Fig. 4A

Fig.4B

Fig. 5

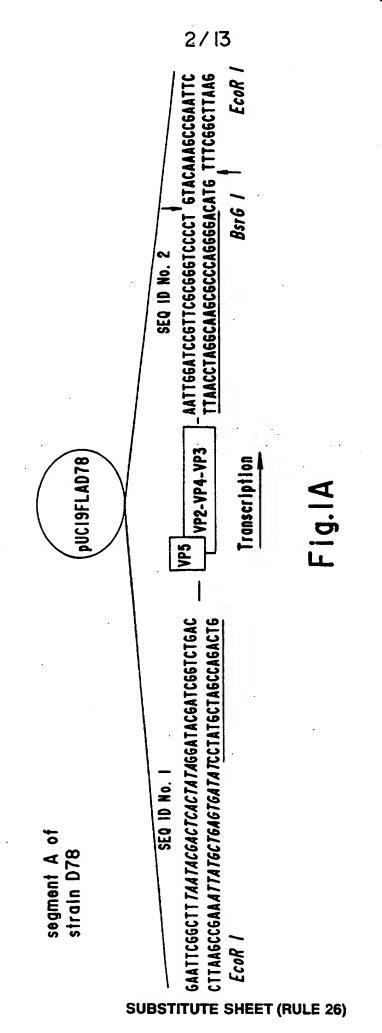
Fig. 5A

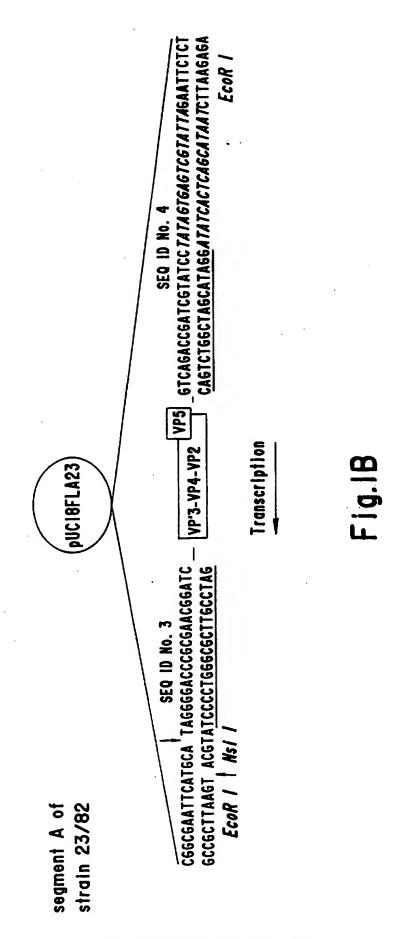
Fig. 5B

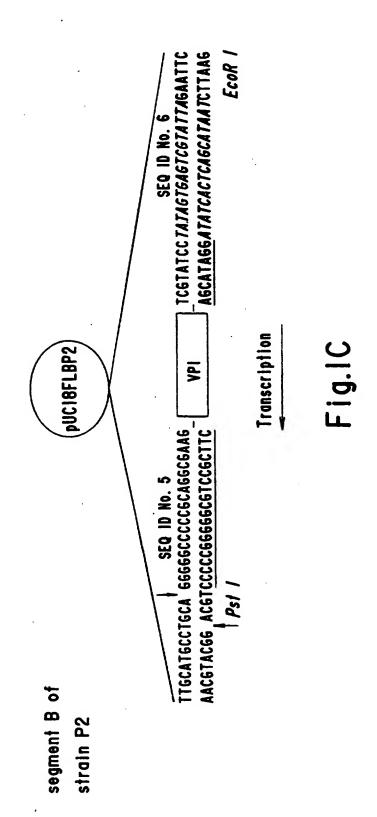
Fig. 6

Fig. 6A

Fig. 6B







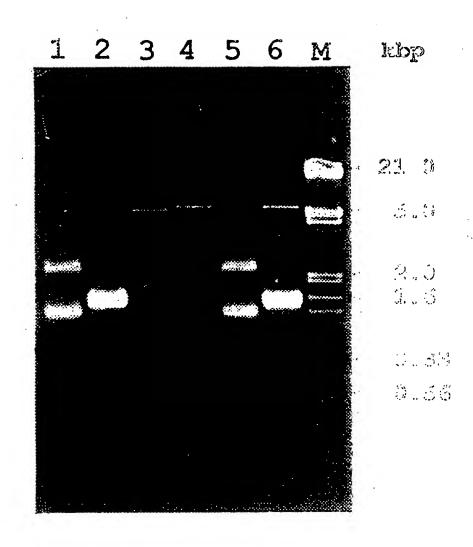


Fig. 2

Fig.3A

	530	540	550	260	570	280
23-82A	GGAAGCCTGAGTGAGTTGACTGACTACAGCTACAACGGGCTGATGTCAGCCACTGCGAAC	SAGTTGACTGA	CTACAGCTAC	AACGGCTGA	TETCAGCCACT	GCGAAC
SEQ ID NO. / 23A/P2B	GGAAGCCTGAGTGAGTTGACTGACTACAGCTACAACGGGCTGATGTCAGCCACTGCGAAC	SAGTIGACTGA	CTACAGCTAC	AACGGCTGA	TETCAGCCACT	GCGAAC
SEQ ID No. 8 P2A	GGAAGCCTGAGTGAACTGACAGATGTTAGCTACAATGGGTTGATGTCTGCAACAGCCAAC	SAACTGACAGATG	r GTTAGCTAC	AAT666TT6/	CTACAATGGGTTGATGTCTGCAACAGCCAAC	
SEQ ID No. 9	230	240	220	260	570	280
	590	009	910	620	630	640
23-82A SEQ ID No. 7	ATCAACGACAAGATCGGGAACGTTCTAGTTGGAGAGGGGTGACTGTTCTCAGTCTACCG	ATCGGGAACGT	TCTAGTTGGA	GAAGGGGTG	ACTETTCTCAG	CTACCE
23A/P2B SEQ 1D No. 8	ATCAACGACAAGATCGGGAACGTTCTAGTTGGAGAAGGGGTGACTGTTCTCAGTCTACCG	ATC666AACGT	TCTAGTTGGA	GAAGGGTG	ACTETTCTCAGI	CTACCE
P2A SEQ ID No. 9	ATCACGACAAATTGGGAACGTCCTAGTAGGGGAAGGGGTCACCGTCCTCAGCTTACCC 590 600 610 620 620	ATTGGGAACGT 600	CCTAGTAGGG	6AA6666TC	ACCETCCTCAGG 630	TTACCC 640

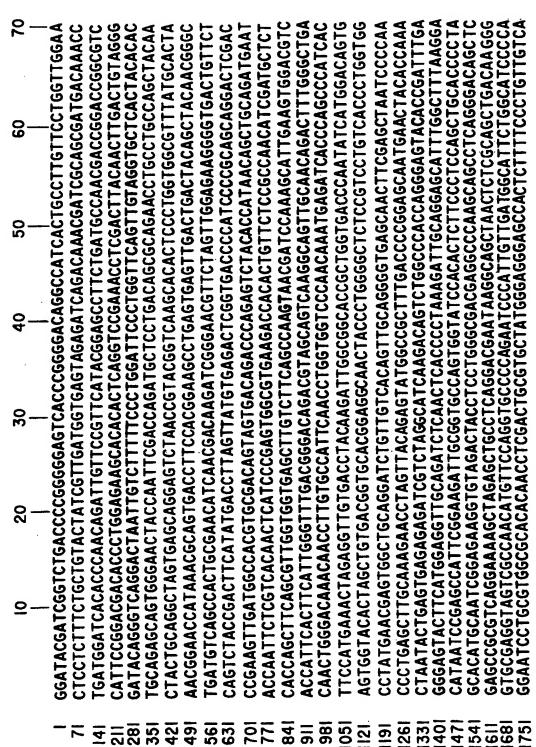
Segment A

Fig.3

	130	140	20		150	<u>&amp;</u>
23-828 SEO ID NO 10	TTTTCAATAGE	TTTCAATAGTCCACAGGCGCAACGAGATCTCAGCAGCGTTCGGCATAAAGCCTACTG	AACGAAGATC	TCAGCAGCGT	TCGGCATAAA	GCCTACTG
23A/P2B	TTTTCAACAGTCCACAGGCGCGCACGATCTCAGCAGGCGTTCGGCATAAAGCCTACTG	TTTCAACAGTCCACAGGGGGGAAGCACGATCTCAGCAGCGTTCGGCATAAAGCCTACT	AGCACGATC	TCAGCAGCET	TCGGCATAAA	GCCTACTG
SEQ 1D NO. 11 P28 SEQ 1D No. 12	TTTCAACAGT	TTTCAACAGTCCACAGGGGGAAGCATCTCAGCATCGGGTTCGGCATAAAGCCTACTG	AGCACGATC 150	TCAGCAGCGT 160	rceccatada 170	6CCTACT6 180
	061	200	210	220	230	240
23-82B Seo ID No. 10	CTGGACAAGACGTGGAAGACTCTTGATCCCCAAAGTCTGGGTGCCACCTGAGGATCCGC	TEGACAAGACGTGGAAGAACTCTTGATCCCCAAAGTCTGGGTGCCACCTGAGGATCCG	TIGATOCCC	AAAGTCT666	FECCACCTEA	66ATCC6C
23A/P2B	CT66ACAA6AC6T66AA6AACTCTT6ATCCCTAAA6TTT666T6CCACCT6A66ATCC6C	STGGAAGAACT(	CTTGATCCCT	AAAGTTT666	TECCACCTEA	GEATCCEC
SEU 10 NO. 11 P2B	CTGGACAAGACGTGGAAGACTCTTGATCCCTAAAGTTTGGGTGCCACCTGAGGATCCGC	CTGGACAAGACGTGGAAGACTCTTGATCCCTAAAGTTTGGGTGCCACCTGAGGATCCG	CTTGATCCCT	AAAGTTT666	TECCACCTEA	66ATCCGC
SEO ID No. 12	190	200	210	220	230	240

Segment B

# Fig.4A



TCCATCTAGCCCTGGCTGCCTCCGAGTTCAAAGAGACCCCAGAACTCGAAGACGCTGTGCGCGCAATGG **GAGGCCCCACACCAGAGAGGCACAGAGGGAAAAGACACACGGATCTCCAAGAAGATGGAAACAATGG** CATCTACTTCGCGACACCGGAATGGGTGGCTCTCAACGGGCACCGAGGCCCAAGCCCCGGCCAACTCAAG ACTEGCAAAACACAAGAGAAATACCAGAGCCCAATGAGGACTACCCAGACTATGTGCACGCGGGAGAGA SAGCAGATGAAGGACCTGCTCCTGACTGCGATGGAGGTGAAGCATCGCAATCCCAGGCGGGCTCCACCAA **AGCCAAAGCCAAAACCCAATGCTCCATCACAGAGCCCCCTGGACGGCTGGGCCGCTGGATCAGGACGGT <b>ATECCECTECAAATECCEACCCATTETTCCECTCAECTCTCCAEGTCTTCATETGETTGGAAGAAACGE** SATTGTGACCGACATGGCTAACTTCGCCCTCAGCGACCCAAACGCGCATAGGATGAAAAACTTCCTAGCA **AACGCACCCCAGGCTGGAAGCAAGTCGCAGAGGGCCAAGTATGGCACGGCAGGCTACGGAGTGGAGGCTC ACCACCCC AGGCCTTCATAGACGAGGTCGCCAGGGTCTATGAAATCAACCATGGGCGTGGTCCAAACCAG** SECTATECCCCAGACGGAGTACTGCCTCTGGAGACCGGGAGAGTACACCGTTGTCCCAATTGATGATG **SCCGGTTG GCGTCAGAAGAACAGATCCTACGGGCAGCCACGTCGATCTACGGGGCTCCAGGACAGGCTGA** CTCCGACG AGGACTTGGAGTGAGGCTCCTGGGAGTCTCCCGACACTACCCGCGCGGGTGT GGACACCAAT **GTGGGACGATAGCATAATGCTGTCGCAGGACCCCATACCTCCAATCATAGGGAACAGCGGCAACCTAGC** CATAGCATACATGGATGTCTTCAGGCCCAAGGTCCCCATCCACGTGGCTATGACAGGGGCCCTCAATGCC CGCGGTGAGATCGAGAGTGTTACGTTCCGVAGCACCAAACTCGCCACAGCCCACCGACTTGGCATGAAGT **AGCTGGTCCTGGAGCCTATGACATTAATACAGGACCTAACTGGGCAACGTTCGTCAAACGTTCCCTCA** SCGAGAGGACCTCCAGCCTCCATCCCAACGGGGATCCTTCATTCGAACTCTCTGGCCATAGAGTCTAT TACGACACTCGAGGATGAGCTGACCCCCAAGGCACTGAACAGCAAAATGTTTGCTGTCATTGAAGGTGT rcgg ccttctaccatcccaaattggatccgttcgcgggtccct 2381 2451 2521 2591 2801 2941 301 3081 2241 2661 2731 2871 2031 201 231 2171

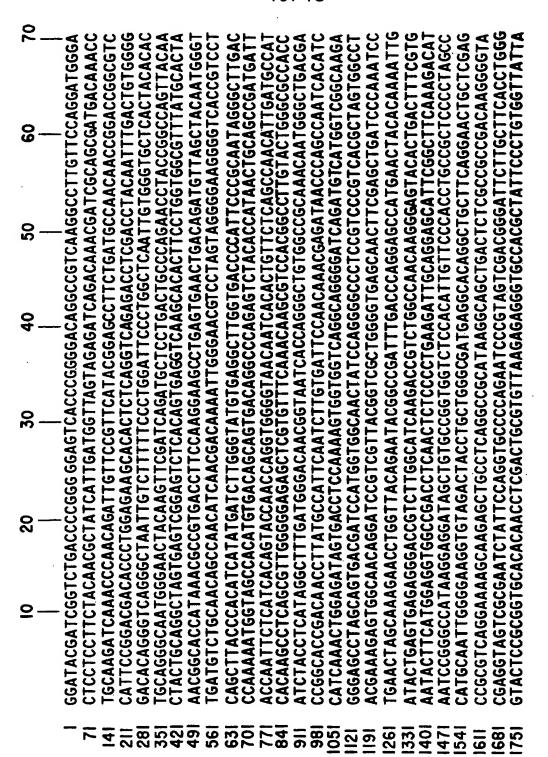
Total number of bases is: 3264. DNA sequence composition: 834 A; 942 C; 853

635

ပ်

Sequence name: 23-82A (SEQ ID NOS: 31 and 33)

Fig.4B



# 11/13

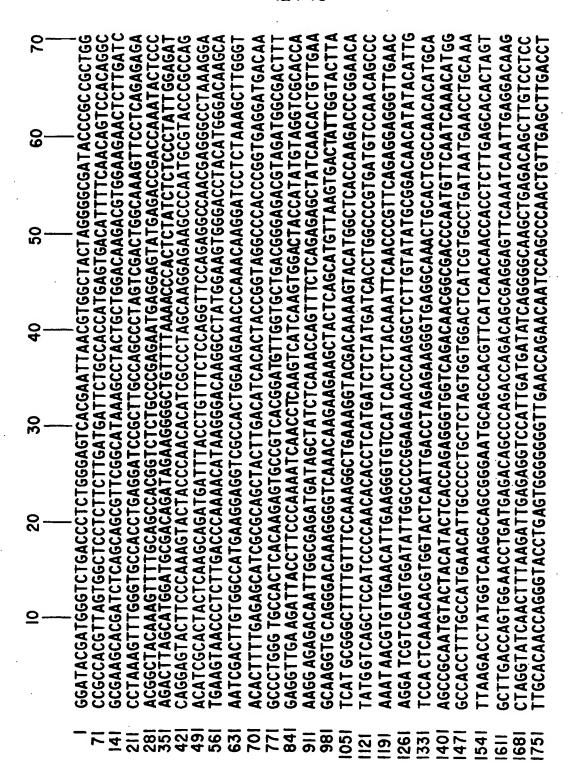
SGACAGTGGAAGACGCCATGACAC CCAAAGCATTGAACAGCAAAATGTTTGCTGTCATTGAAGGCGTGCG **TGGTCCCGGAGCATTCGATGTAAACACCGGGCCCAACTGGGCAACGTTCATCAAACGTTTCCCTCACAA CCACGCCACTGGGACAGGCTCCC.CTACCTCAACCTACCATACCTTCCACCCAATGCAGGACGCCAGTAC ACCTTGCCATGGCTGCATCAGAGTTCAAAGAGACCCCGGAACTCGAGAGTGCCGTCAGAGCAATGGAAG GTGACTGACATGGCCAACTTCGCACTCAGCGACCCGAACGCCCATCGGATGCGAAATTTTCTTGCAAAC ACCCCAAGCTTTCATAGACGAAGTTGCCAAAGTCTATGAAATCAACCATGGACGTGGCCCAAACCAAGAA 286ATGAAAGATCTGCTCTTGACTGCGATGGAGATGAAGCATCGCAATCCCAGGCGGGCTCTACCAAAGC** CAAGCCAAAACCCAATGCTCCAACACAGAGACCCCTGGTCGGCTGGGCCGCTGGATCAGGACCGTCTC **AGAAGACCTCCAACCTCCATCTCAAAGAGGATCCTTCATACGAACTCTCTGGACACAGAGTCTATGGA 3GCGAGATTGAGAAAGTAAGCTTTAGAAGCACCAAGCTCGCCACTGCACACCGACTTGGCCTTAGGTTGG AGCAGCCAACGTGGACCCACTATTCCAATCTGCACTCAGTGTGTTCATGTGGCTGGAAGAGAGATGGGAT ICACCACAAGCAGCAGCAAGTCGCAAAGGGCCAAGTACGGGACAGCAGGCTACGGAGTGGAGGCTCGGG ATGCTCCAGATGGGGTACTTCCACTGGAGACTGGGAGAGACTACACCGTTGTCCCAATAGATGATGTCT SCCCCACACCACAGAGGAAGCACAGAGGGAAAAAGACACACGGATCTCAAAGAAGATGGAGACCATGGGCAT** GATGAGGACCTTGAGTGAGGCTCCTGGGAGTCTCCCGACACCCCCGCGCGGGTGTGGGACACCAATTCG **3GGACGACAGCATTATGCTGTCCAAAGATCCCATACCTCCTATTGTGGGAAACAGTGGAAATCTAGCCAT** ;TACTTTGCAACACCAGAATGGGTAGCACTCAATGGGCACCGAGGGCCAAGCCCCGGCCAGCTAAAGTAC **<b>4**GCTTACATGGATGTTTCGACCCAAAGTCCCAATCCATGTGGCTATGACGGGAGCCCTCAATGCTTGT **GGTTGGCATCAGAAGAACAAATCCTAAGGGCAGCTACGTCGATCTACGGGGCTCCAGGACAGGCAGAGCC** SCCTTACAACATCCCAAATTGGATCCGTTCGCGGGTCCCCT 2801 2941 2031 2241 2381 2451 2521 2591 2661 2731 2871 301 3081 196 2101 2171 2311 3151 89

Total number of bases is: 3261.

DNA sequence composition: 873 A; 909 C; 847 G; 632 T; 0 OTHER;

Sequence name: D78F (SEQ ID NOS: 27 and 29)

Fig.5E



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TGTTGGGCTCCACCTGCCCGCCAAGAGAGCCACCGGTGTCCAGGCCGCTCTTCTCGGAGCAGGAACGAG CAGACCAATGGGGATGGAGGCCCCAACACGGTCCAAGAACGCCGTGAAAATGGCCAAACGGCGGCAACGC :TCGTCCTTCTAGCCACAGCAAGAAGCCGTCTGCAAGATGCAGTTAAGGCCAAGGCAGAAGCCGAGAAAC STTCAGTCGACTTCCGTGTACACCCCCAAGTACCCAGAAGTCAAGAACCCACAGACCGCCTCCAACCCCG CAAAAGGAGAGCCGCTAACAGCCATGATGGGAACCACTCAAGAAGAGGACACTAATCCCAGACCCCGTAT GAGAAAGCCGACATCGCCAGCAAGGTCGCCCACTCAGCACTCGTGGAAACAAGCGACGCCCTTGAAGCA **<b>ACTAGGGTGGTCAGCTACATACAGCAAAGATCTCGGGATCTATGTGCCGGTGCTTGACAAGGAACGCCTA** TTTGTTCTGCTGCGTATCCCAAGGGAGTAGAGAACAAGAGTCTCAAGTCCAAAGTCGGGATCGAGCAGG :ATACAAGGTAGTCAGGTATGAGGCGTTGAGGTTGGTAGGTGGTTGGAACTACCCACTCCTGAACAAAGC CTGCAAGAATAACGCAGGCGCCGCTCGGCGCATCTGGAGGCCAAGGGGTTCCCACTCGACGAGTTCCTA **GCCGAGTGGTCTGAGCTGTCAGAGTTCGGTGAGGCCTTCGAAGGCTTCAATATCAAGCTGACCGTAACAT TEAGAGCCTAGCCGAACTGAACAAGCCAGTACCCCCCAAGCCCCCAAATGTCAACAGACCAGTCAACAC** CCACAAGTCCAAGCCAGACGACCCCGATGCAGACTGGTTCGAAAGATCAGAACTCTGTCAGACCTTC1 cccesecttcscctscssssscc 2451 2591 273 2031 2241 2311 2381 **252**1 2661 196 2101 2171

Total number of bases is: 2827.

DNA sequence composition: 796 A; 770 C; 724 G; 537

Sequence name: P2B (SEQ ID No: 25)

- ig.6B

## INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/12955

A. CLASSIFICATION OF SUBJECT MATTER  IPC(6) :Please See Extra Sheet.						
US CL : Please See Extra Sheet.						
According	to International Patent Classification (IPC) or to both	national classification and IPC				
	LDS SEARCHED					
Minimum documentation searched (classification system followed by classification symbols)						
U.S. :	424/184.1, 204.1, 816, 826; 435/71.1, 235.1, 236,	237, 238, 239, 320.1; 536/23.72				
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched						
	data base consulted during the international search (n N-MEDLINE, BIOSIS, CAPLUS, CABA	ame of data base and, where practicabl	e, search terms used)			
C. DOC	CUMENTS CONSIDERED TO BE RELEVANT		·			
Category*	Citation of document, with indication, where a	ppropriate, of the relevant passages	Relevant to claim No.			
х	MUNDT et al. Complete Nucleotid Noncoding Regions of Both Genome S of Infectious Bursal Disease Virus. Vir 10-18, see entire document.	Segments of Different Strains	1-2, 4-20			
X	US 4,530,831 A (LUTTICKEN ET Al see entire document.	L) 23 JULY 1985 (07/23/85),	7, 15-20			
X	US 5,192,539 A (VAN DER MAREL ET AL) 09 MARCH 1993 1-3, 7, 15-20 (09/03/93), see entire document.					
x	MUNDT et al. Identification of a novel viral protein in infectious bursal disease virus-infected cells. Journal of General Virology. 1995, Vol. 76, pages 437-443, see entire document.					
X Further documents are listed in the continuation of Box C. See patent family annex.						
Special categories of cited documents:						
"A" document defining the general state of the art which is not considered the principle or theory underlying the invention						
to be of particular relevance  "X" document of particular relevance; the claimed invention cannot be						
*L* document which may throw doubts on priority claim(s) or which is considered novel or cannot be considered to involve an inventive s when the document is taken alone						
cited to establish the publication date of another citation or other special reason (as specified)  "Y" document of particular relevance; the claimed invention cannot be						
	considered to involve an inventive step when the document is					
	cument published prior to the internstional filing date but later than a priority date claimed	"A" document member of the same paten	t family			
Date of the	actual completion of the international search	Date of mailing of the international se	arch report			
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# INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/12955

	tion). DOCUMENTS CONSIDERED TO BE RELEVANT	<del></del>		
Category*	Citation of document, with indication, where appropriate, of the relevan	nt passages	Relev	ant to claim No
ζ	BAYLISS et al. A comparison of the sequences of segme four infectious bursal disease virus strain and identification variable region in VP2. Journal of General Virology. 199 71, pages 1303-1312, see entire document.	on of a	1-2, 5	-8, 10-13
	MORGAN et al. Sequence of the Small Double-Stranded Genomic Segment of Infectious Bursal Disease Virus and Deduced 90kDa Product. Virology. 1988, Vol. 163, pages see entire document.	l Its	1-20	
	SPIES et al. Nucleotide sequence of infectious bursal dis- genome segment A delineates two major open reading fra Nucleic Acids Research. 1989, Vol. 17, No. 19, page 798 entire document.	ames.	1-20	· · ·
,	WO 91/16925 A1 (UNIVERSITY OF MARYLAND at COLLEGE PARK) 14 NOVEMBER 1991 (14/11/91), see document.	e entire	1-20	
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### INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/12955

A.	CLASSIFICATION	OF	SUBJECT	MATTER:
IP	C (6):			

A61K 39/00, 39/38, 39/12; C12P 21/04; C12N 7/00, 7/01, 7/02, 7/04, 7/06, 7/08, 15/00, 15/09, 15/63, 15/70, 15/74

A. CLASSIFICATION OF SUBJECT MATTER: US CL :

424/184.1, 204.1, 816, 826; 435/71.1, 235.1, 236, 237, 238, 239, 320.1; 536/23.72